

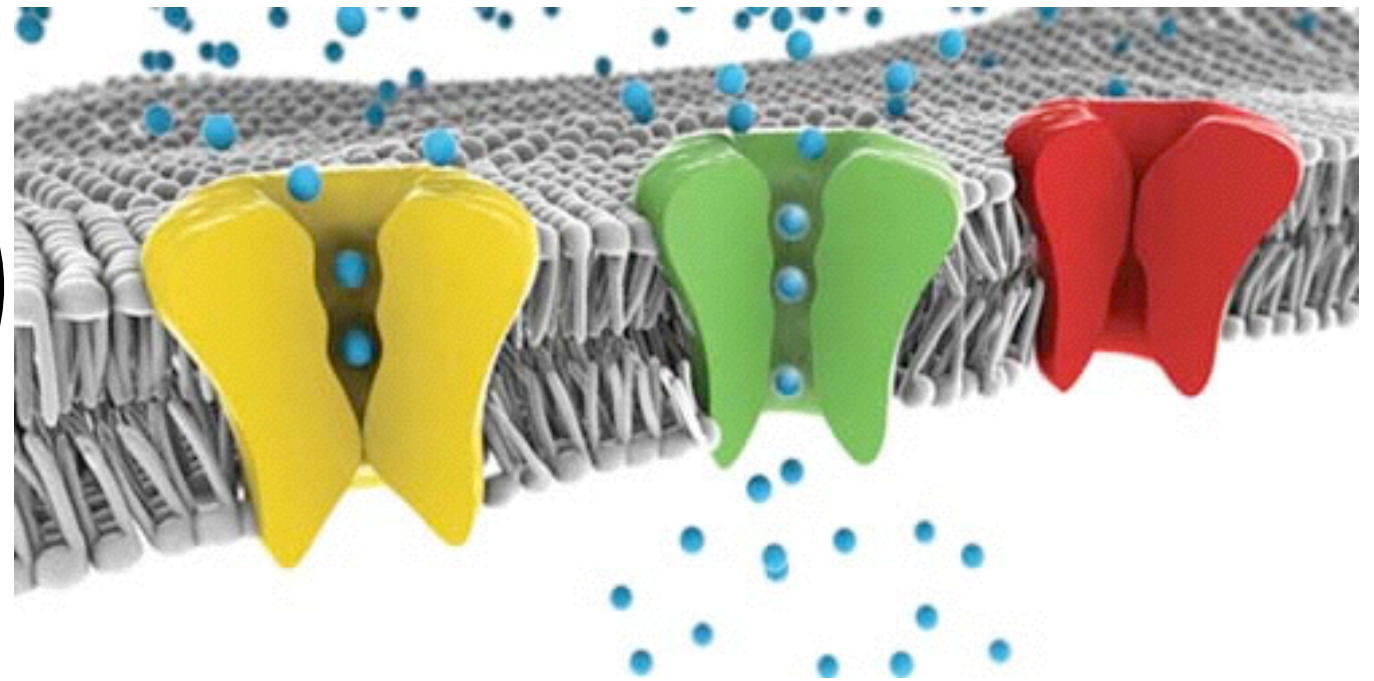
Membranes

18.354 - L11

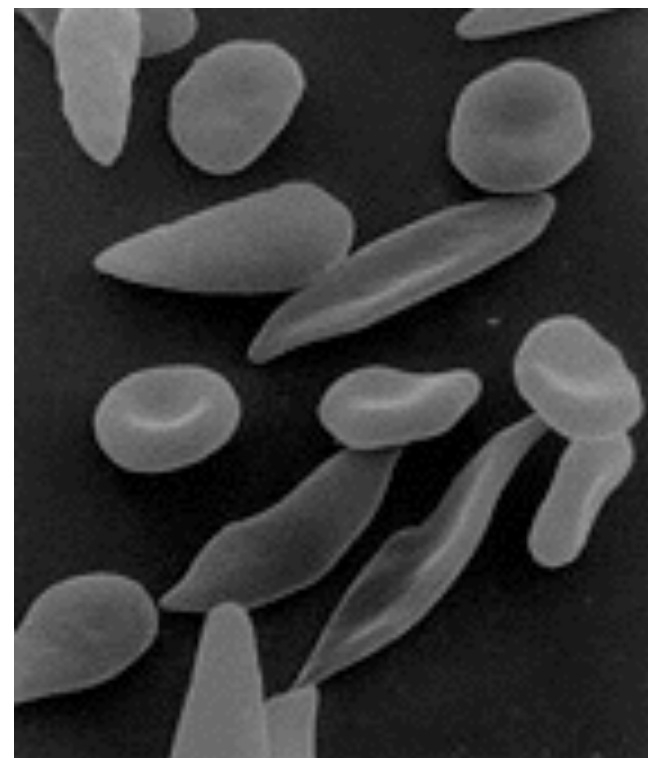
Cell membranes ($D=2$)

Illustration by J.P. Cartailier. Copyright 2007, Symmation LLC.

transport:
stochastic
escape problems



shape:
differential
geometry

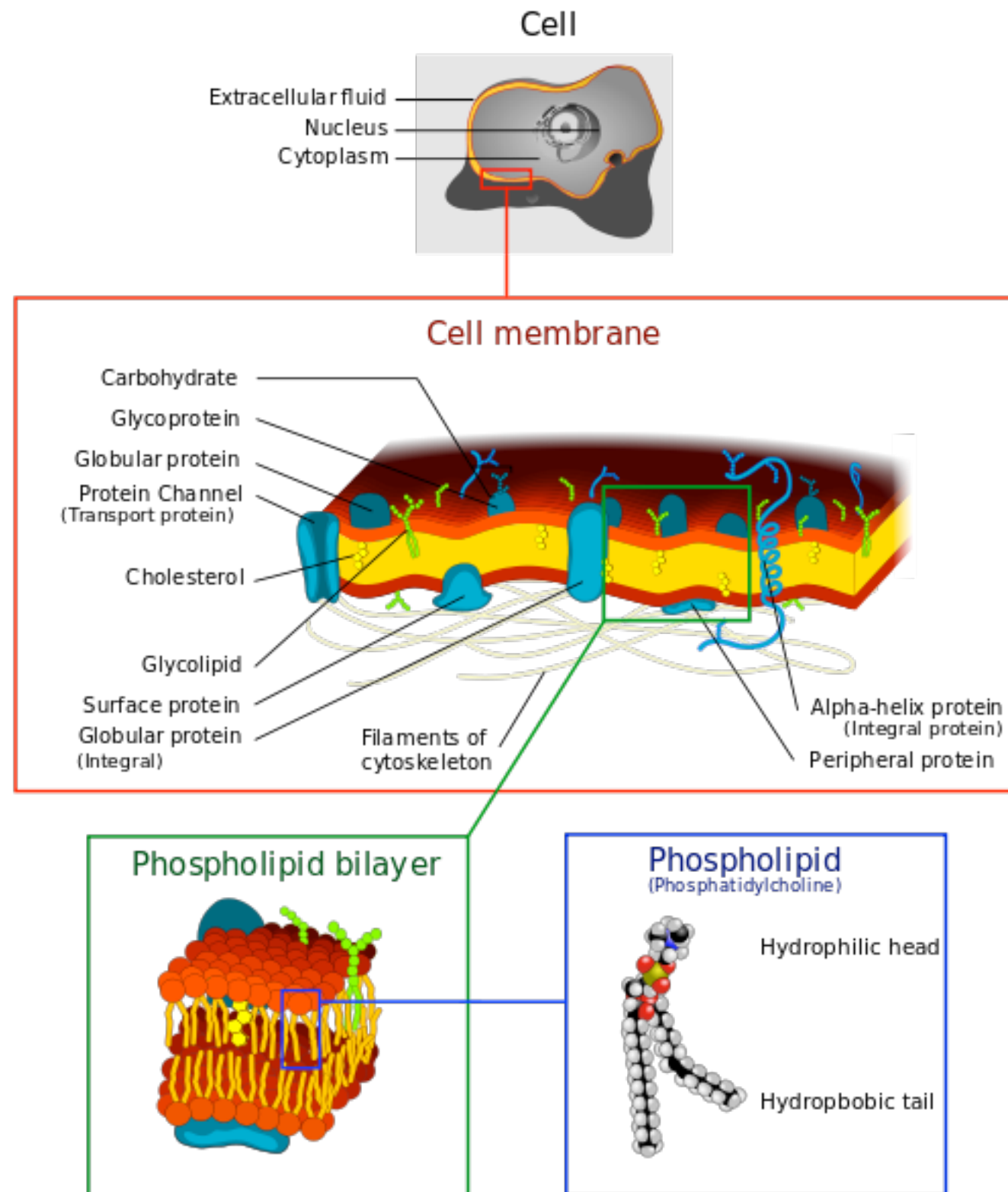


red blood cells
affected by
sickle-cell disease

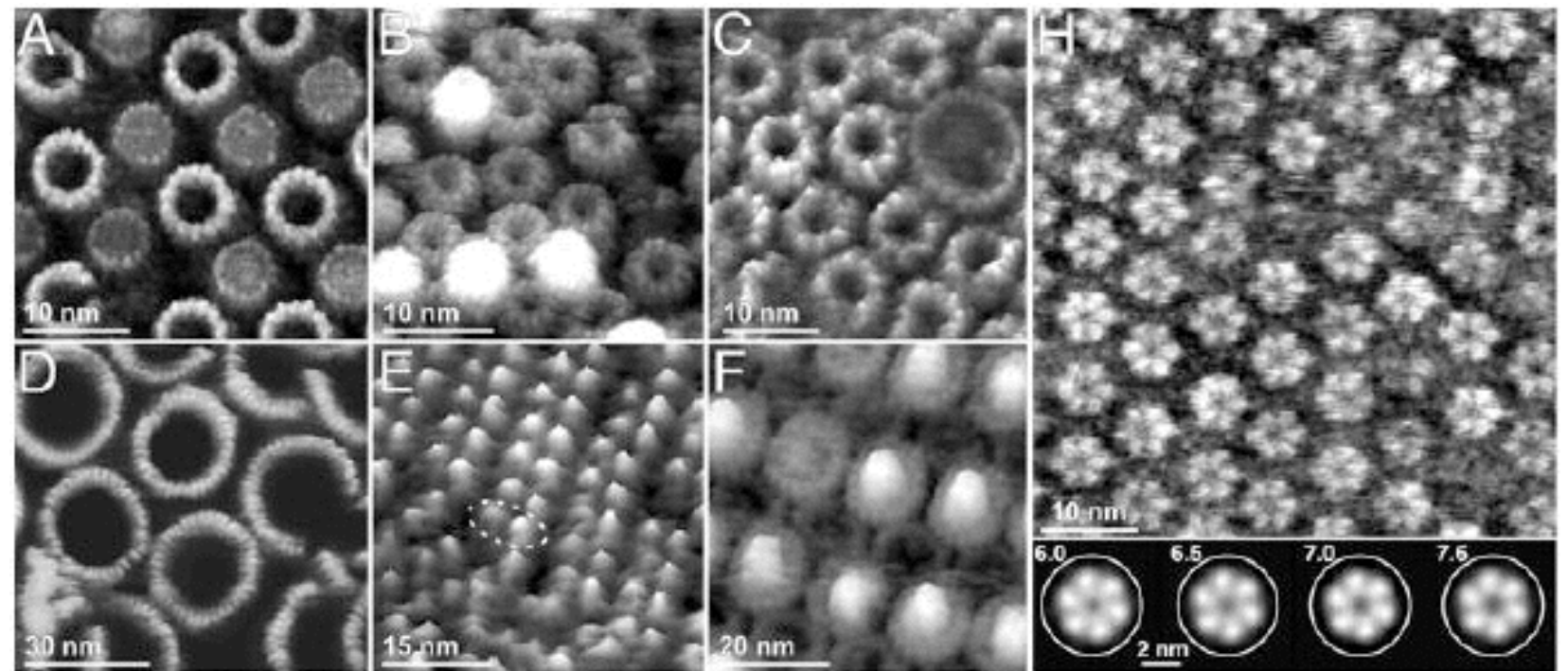
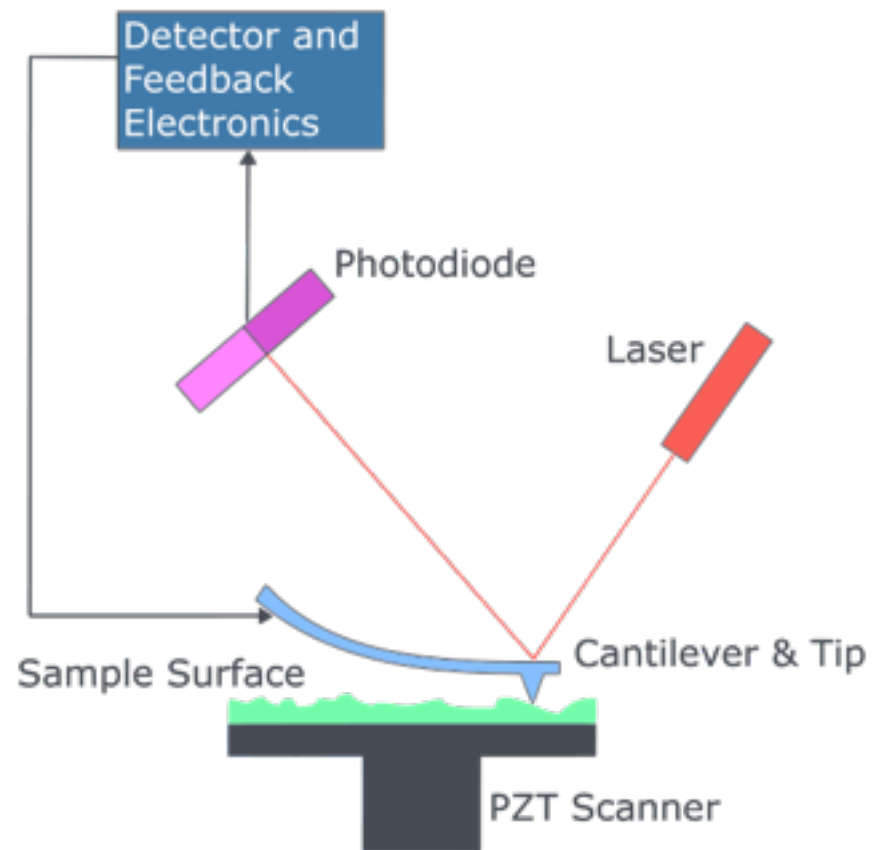
source: wiki

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Cell membranes



AFM

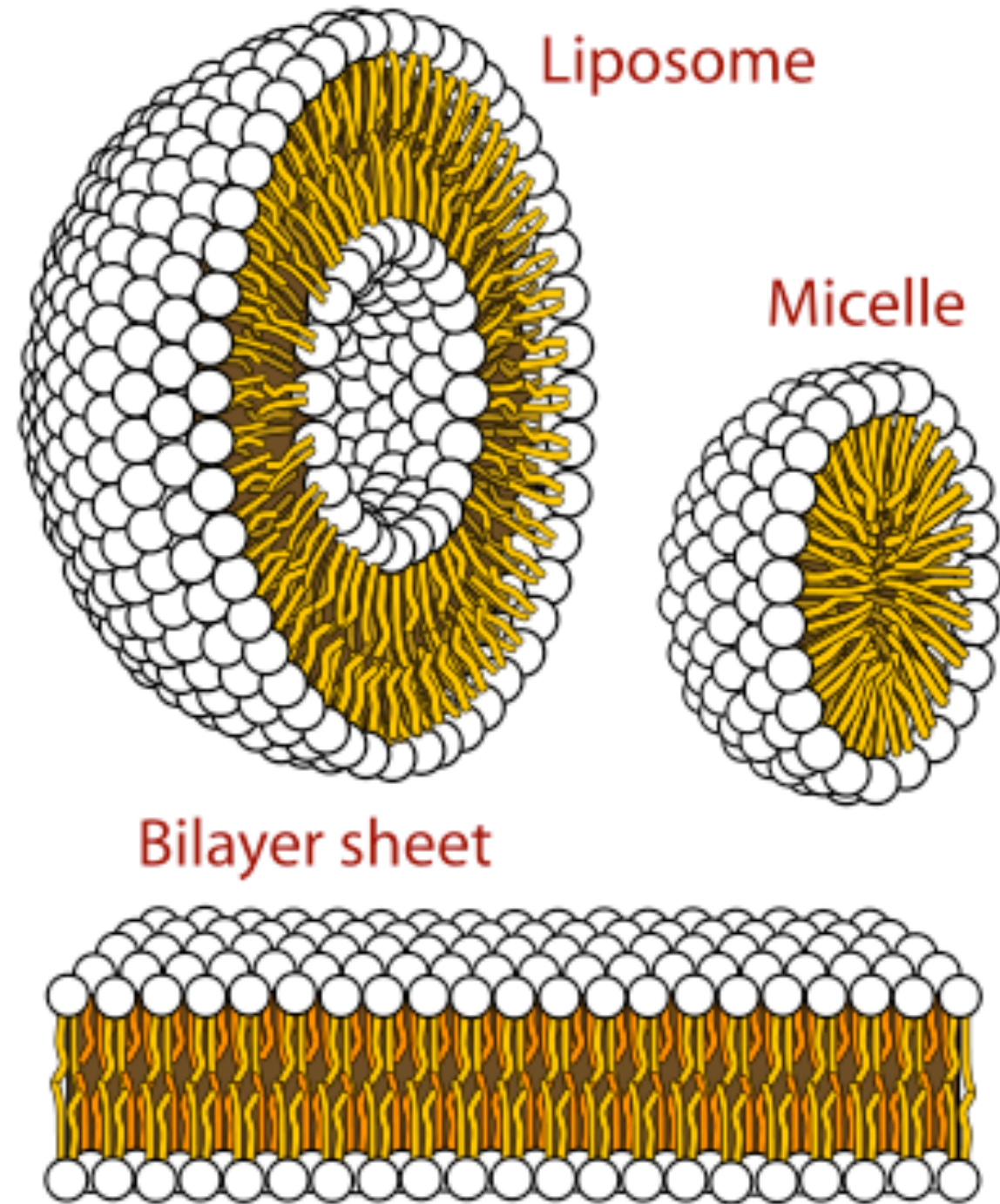
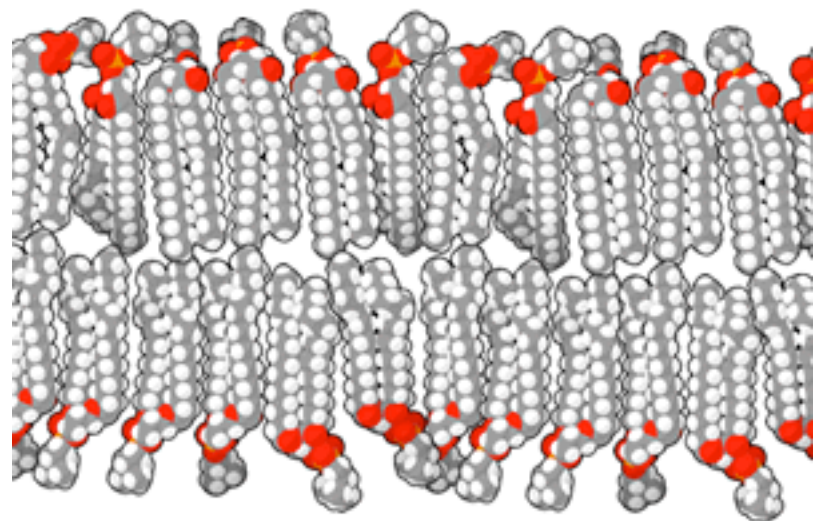


http://www.sbmp-itn.eu/sbmpps/research_method/

source: wiki

Bio-membrane

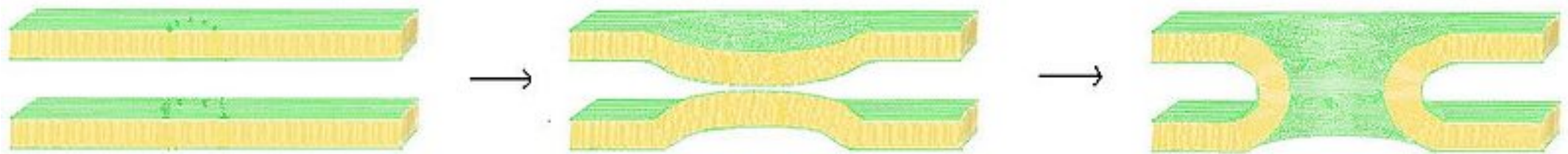
Selective barrier



source: wiki

Morphological changes

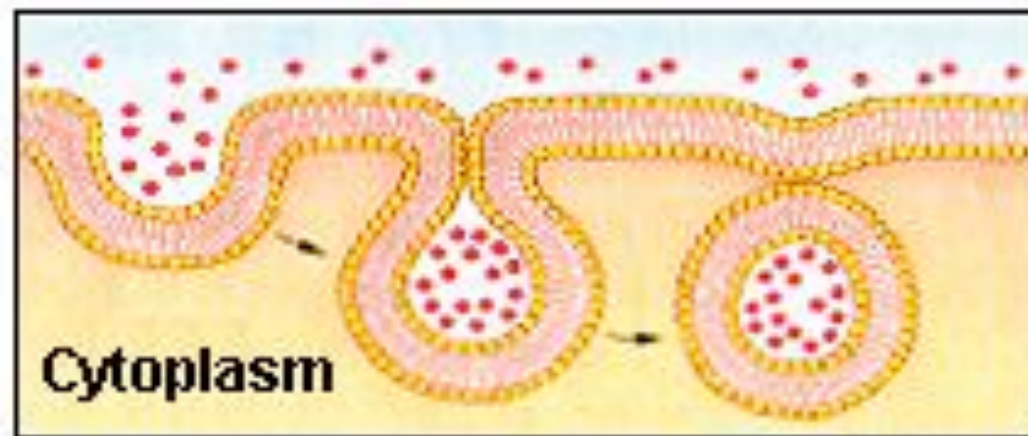
e.g., fusion through stalk-formation



source: wiki

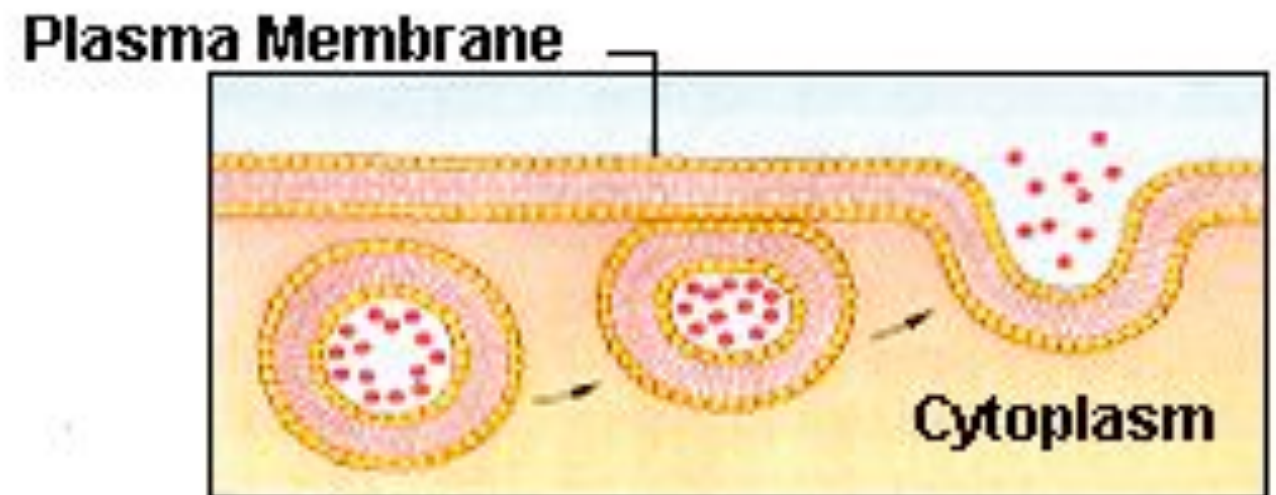
Endo- & Exocytosis

material exchange with environment



Endocytosis

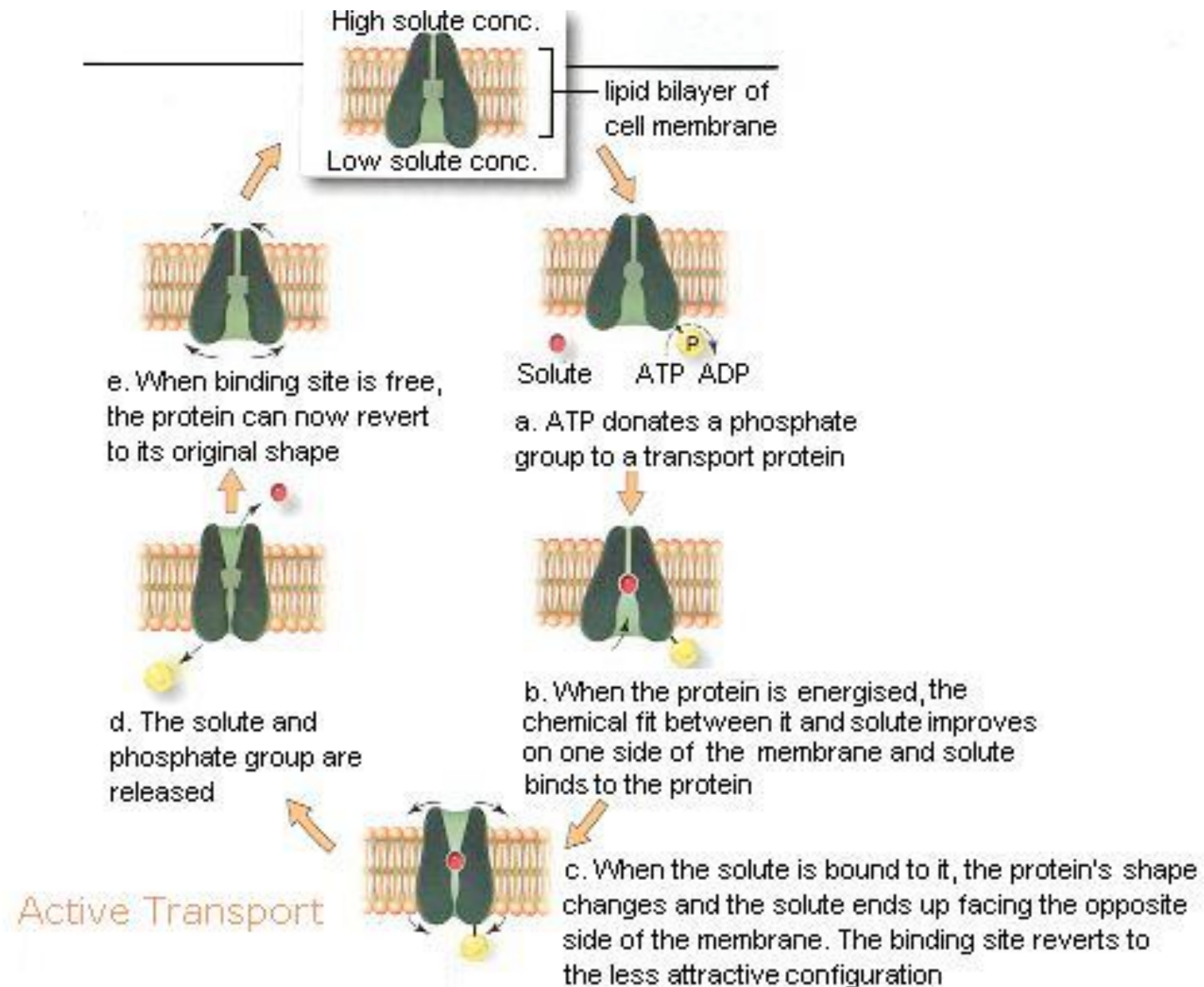
A bit of plasma membrane balloons inward beneath water and solutes outside, then pinches off as an endocytic vesicle that moves into the cytoplasm



Exocytosis

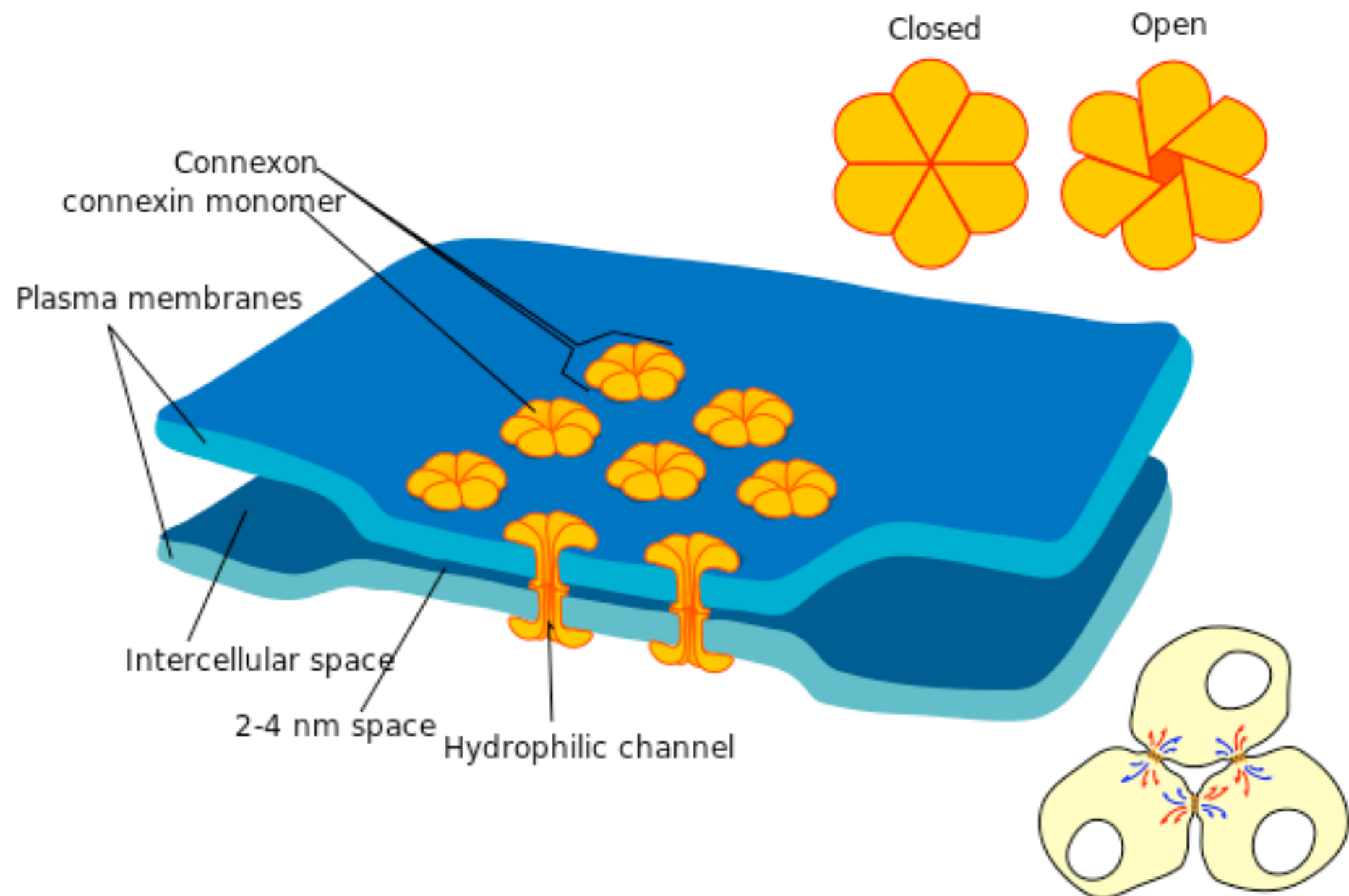
Cells release substances when an exocytic vesicle's membrane fuses with the plasma membrane

Active transport



Intercellular gap junctions

exchange of molecules and ions between **animal** cells

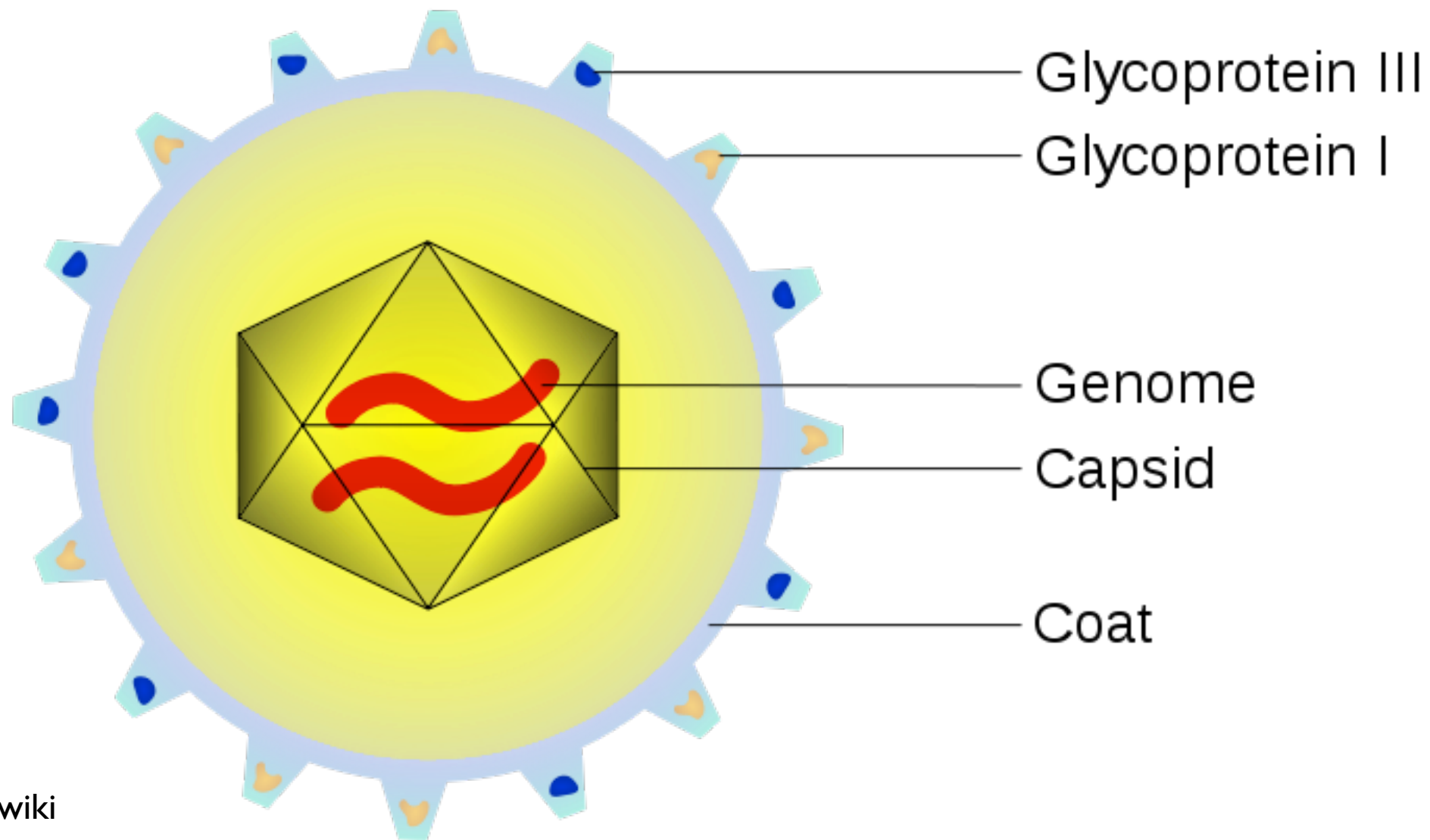


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Virus envelop

Scheme of a CMV virus

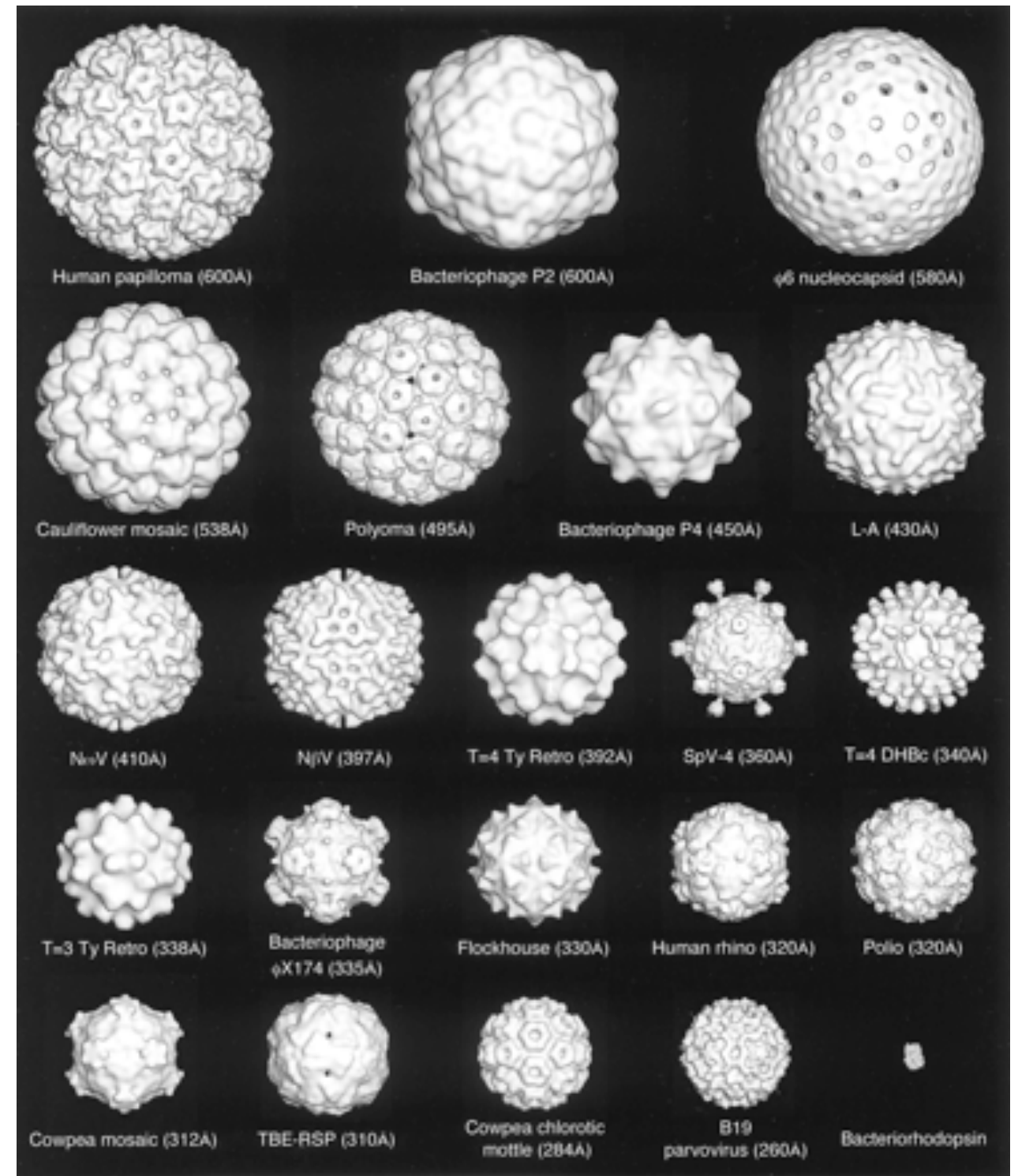
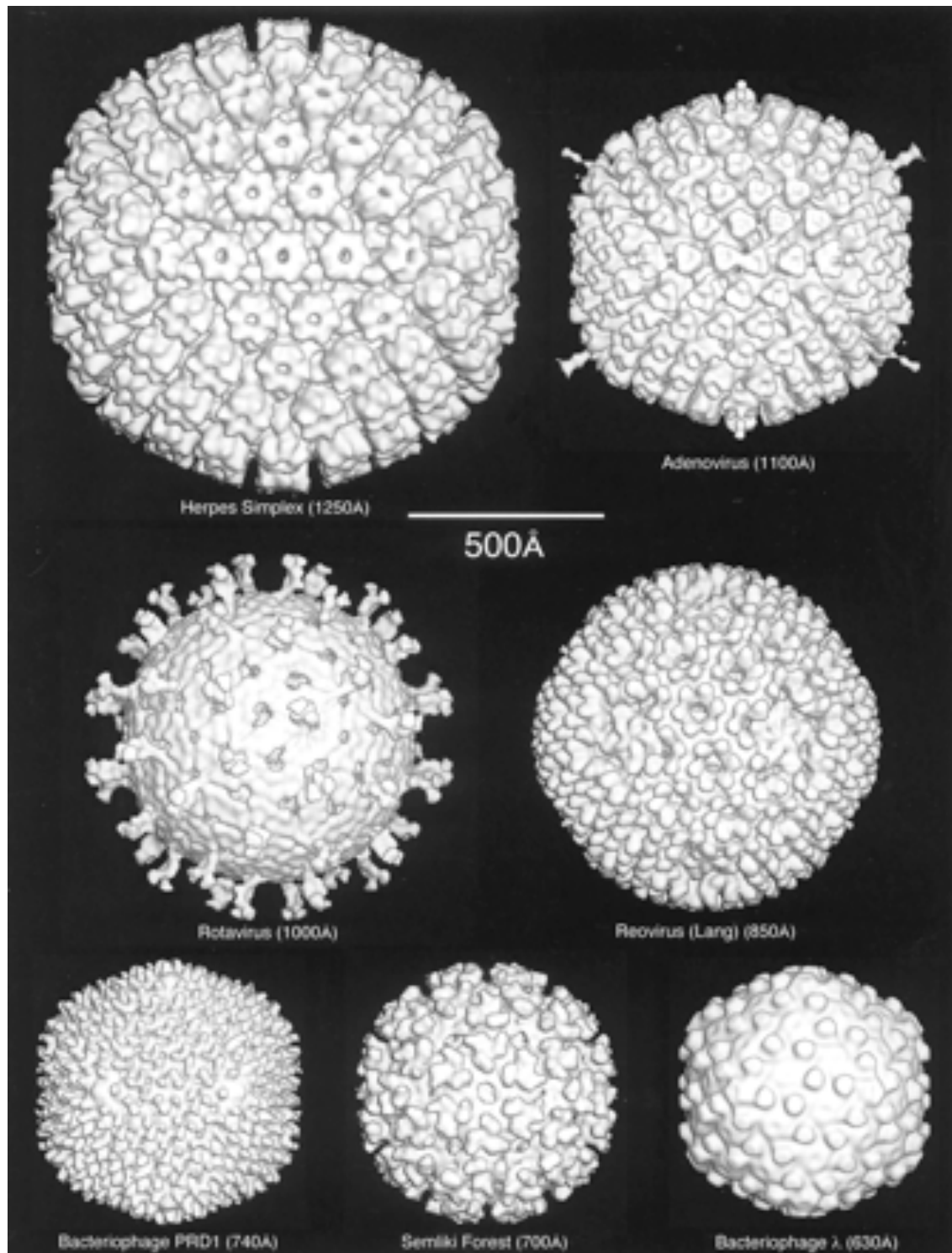


source: wiki

envelop fuses with host membrane

Capsid

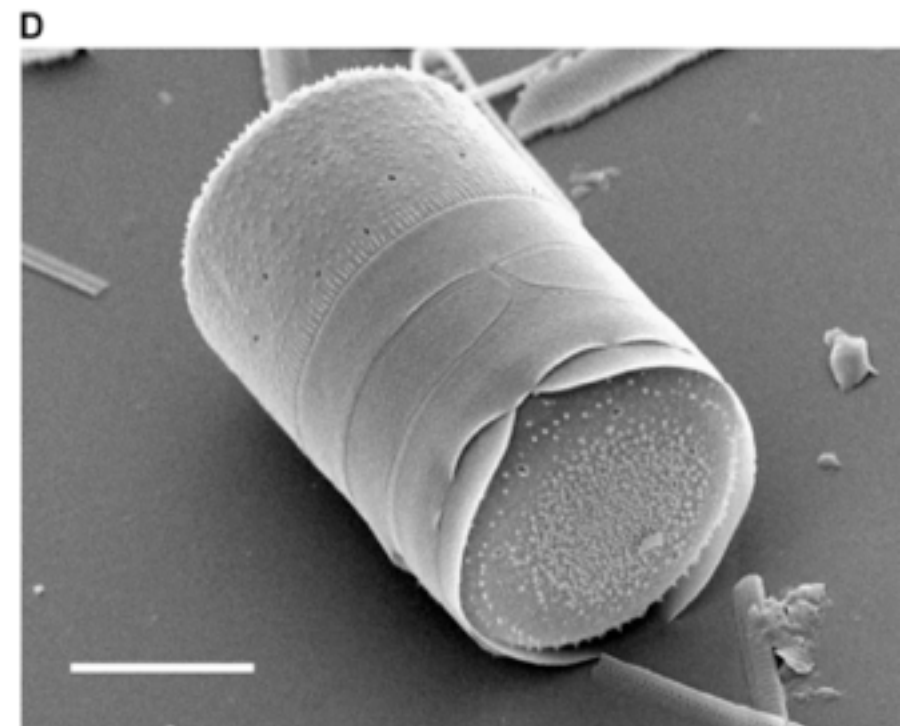
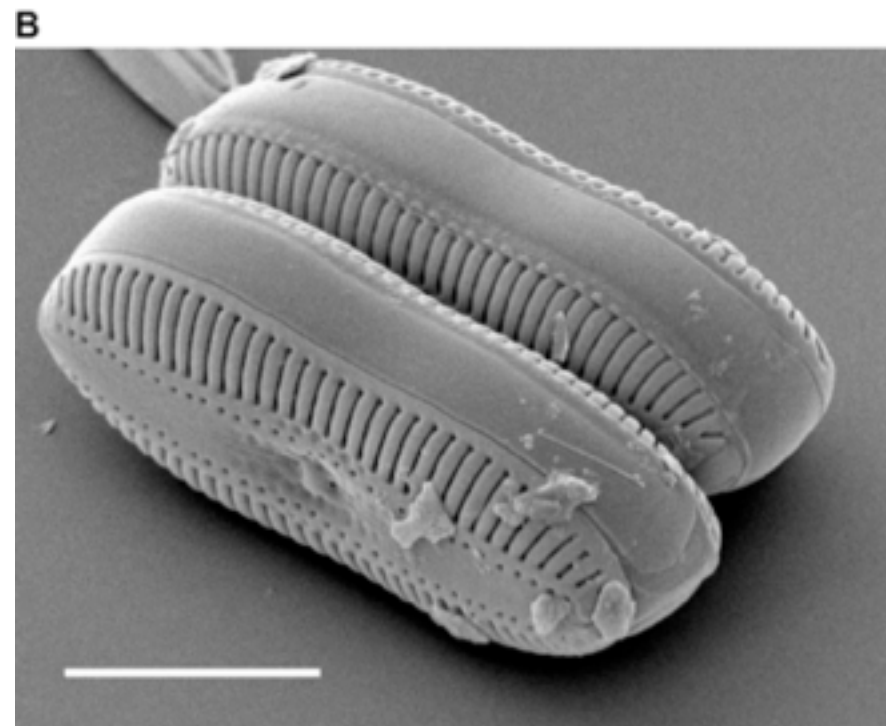
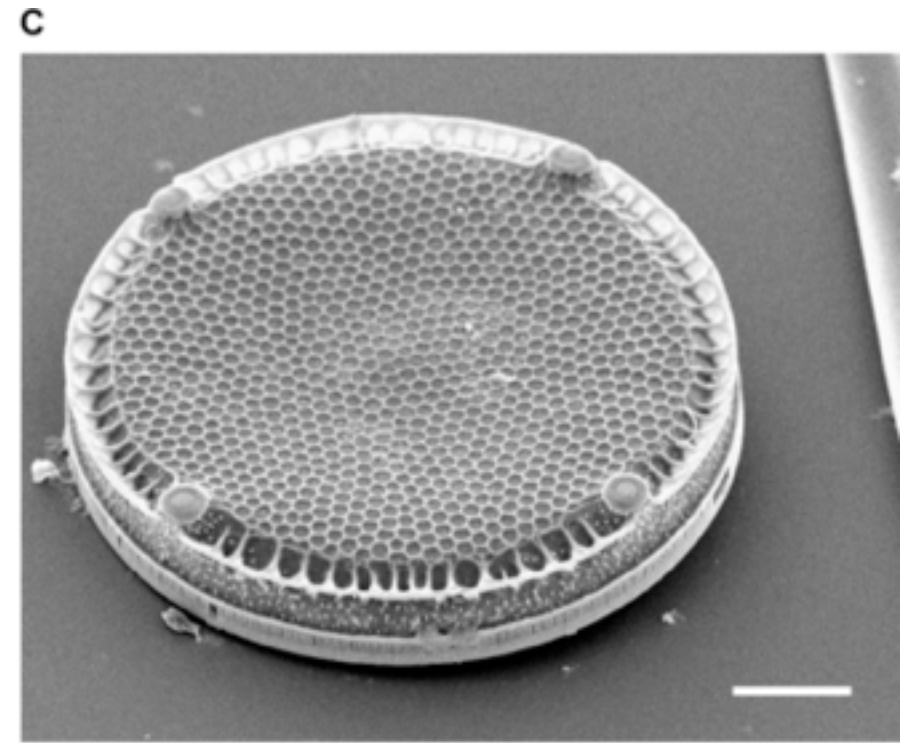
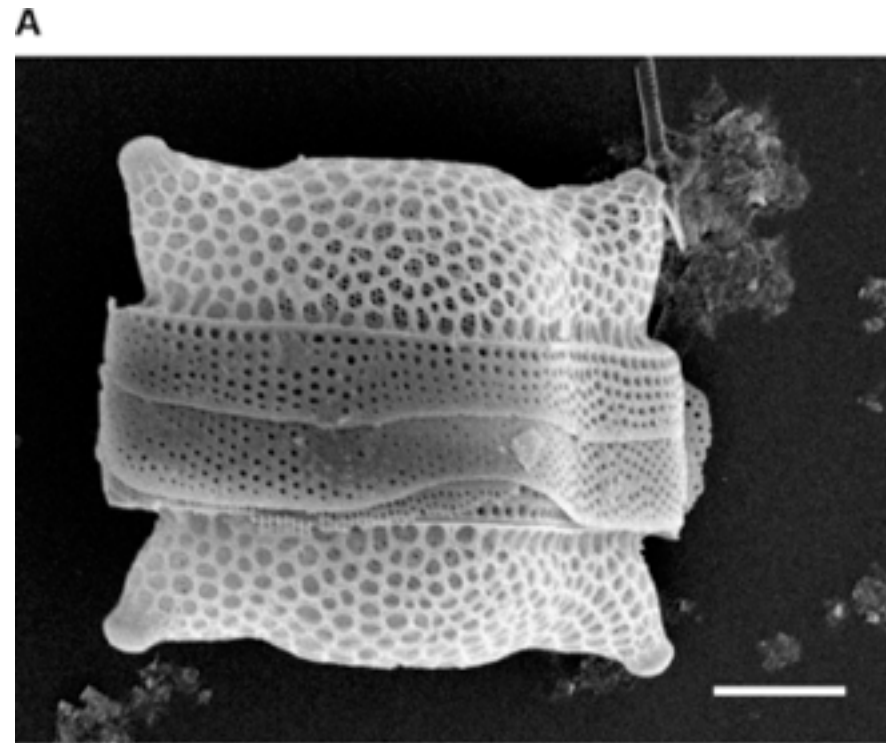
cryo-EM



Baker et al (1999) MMBR

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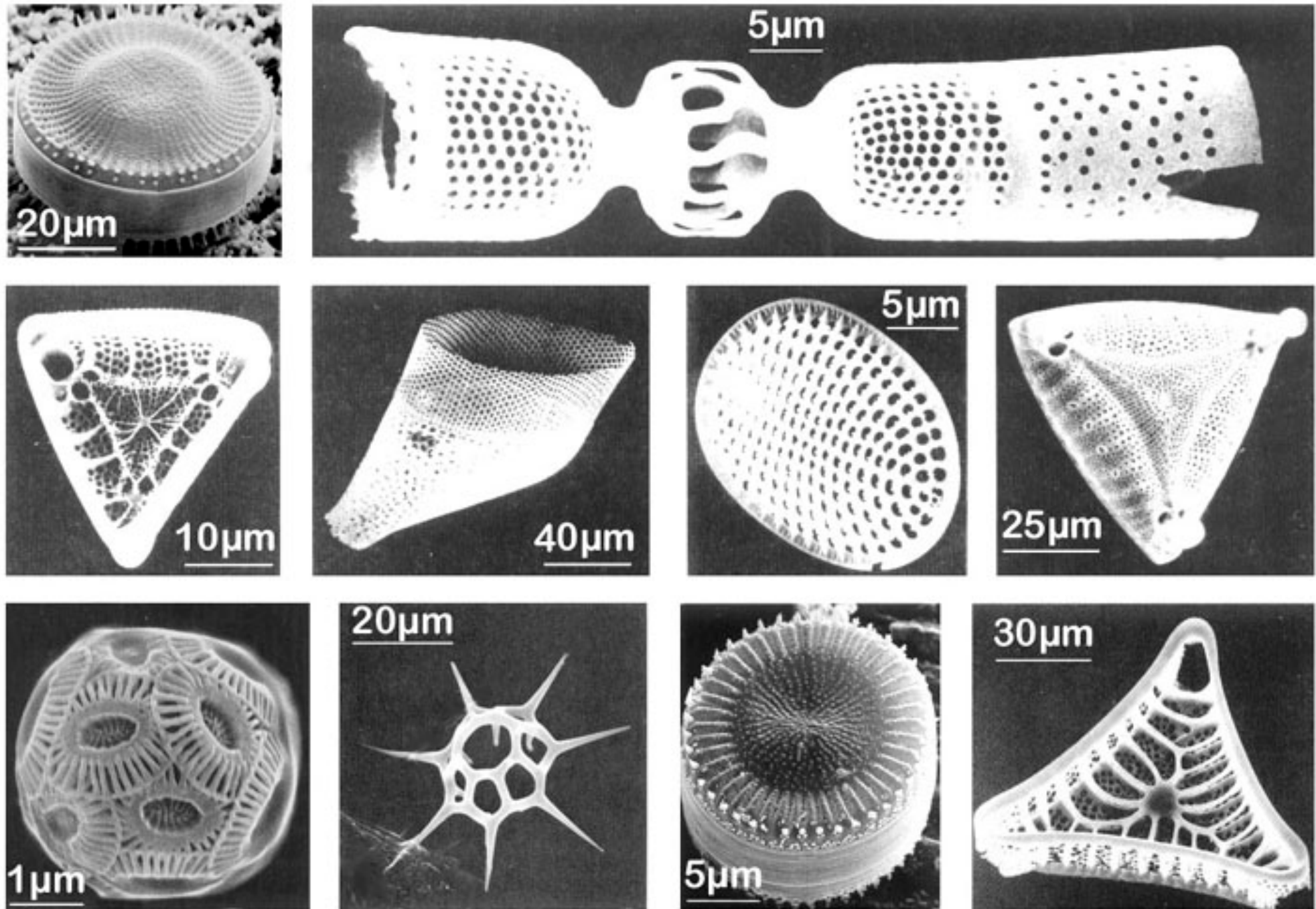
Diatoms (algae)



source: wiki

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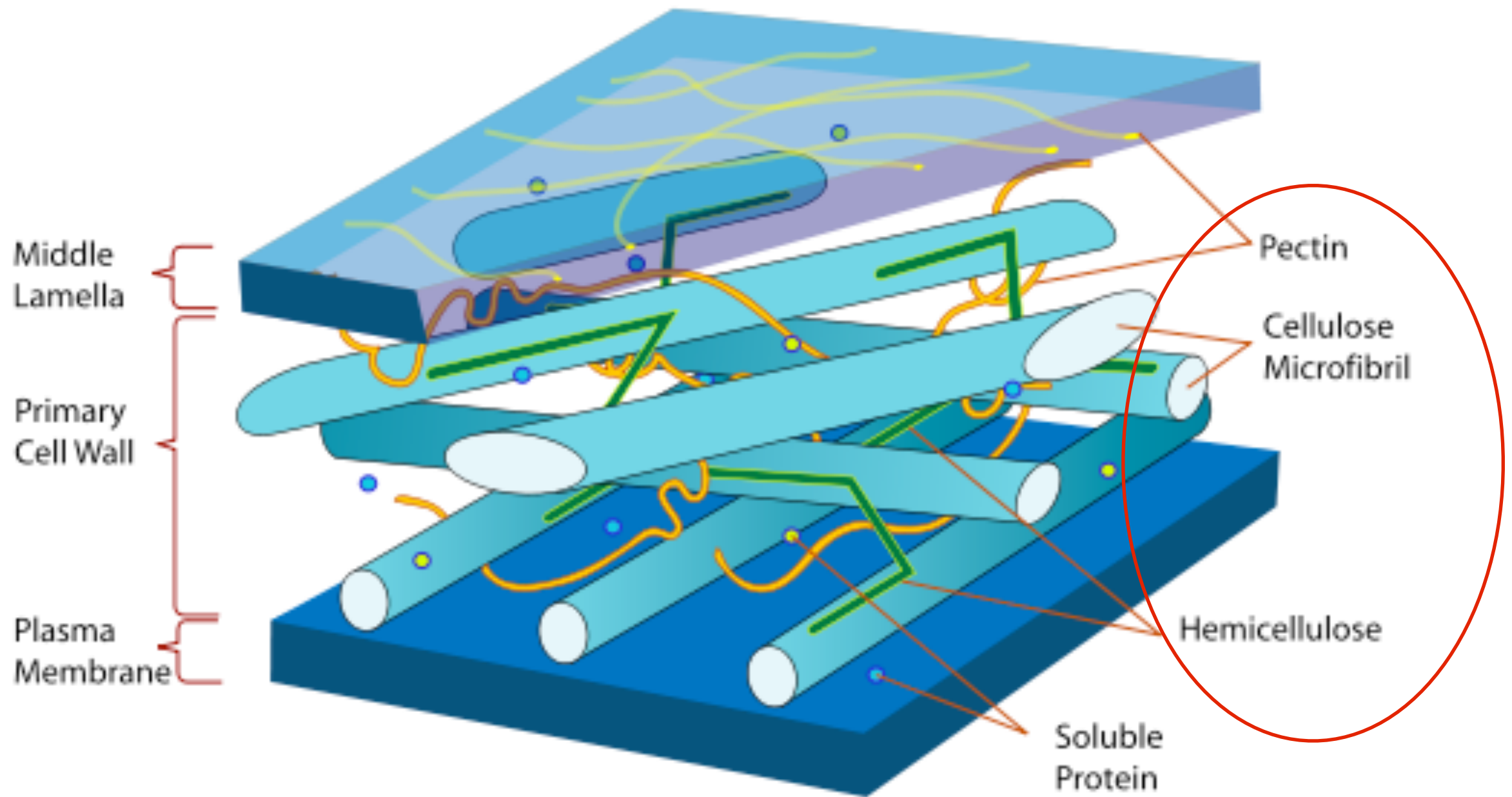
More planktonic diatoms

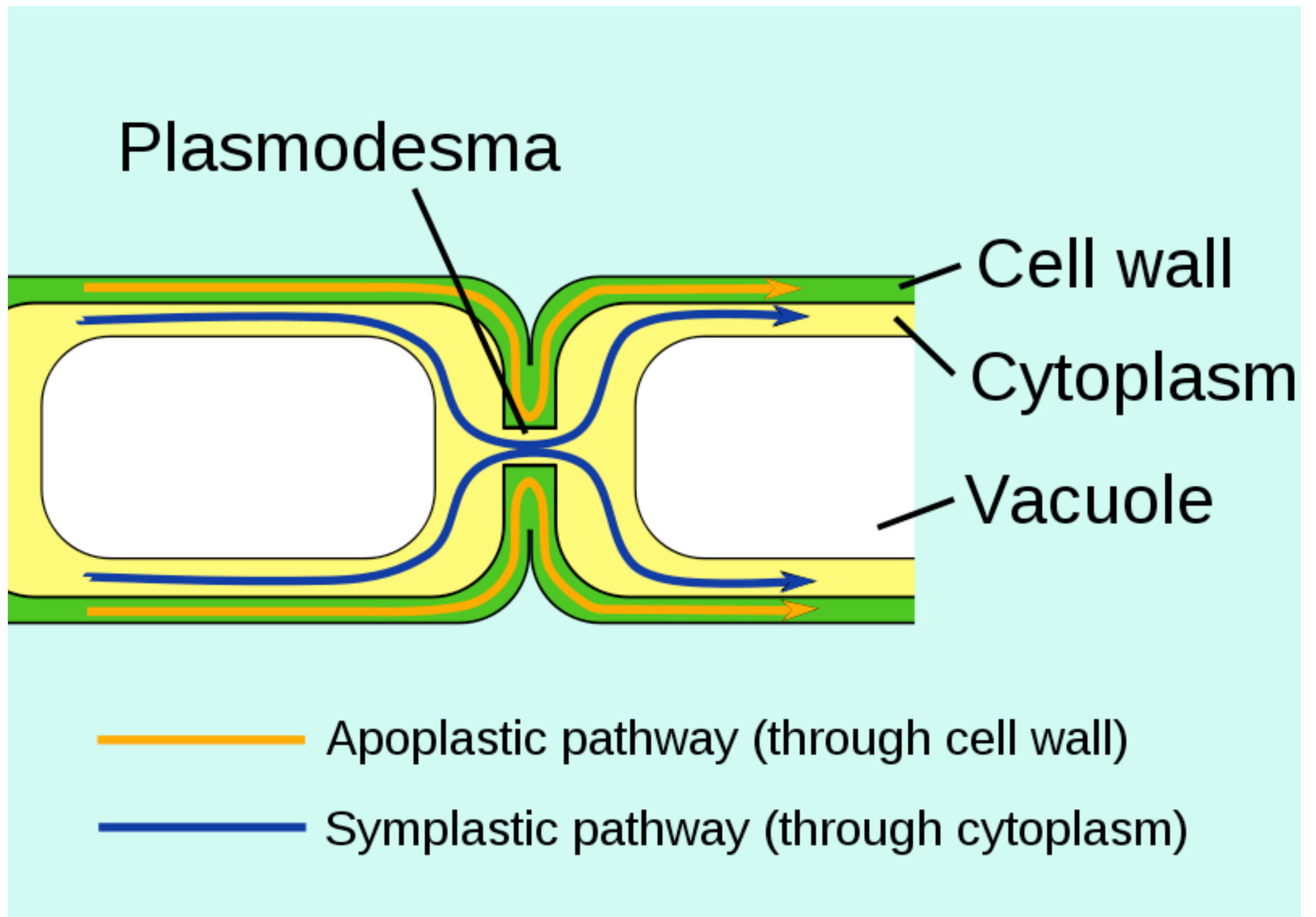


Selection of planktonic diatoms
(not representative for the mediterranean)

Plants

unlike **animal cells**, every **plant cell** is surrounded by a **polysaccharide cell wall**





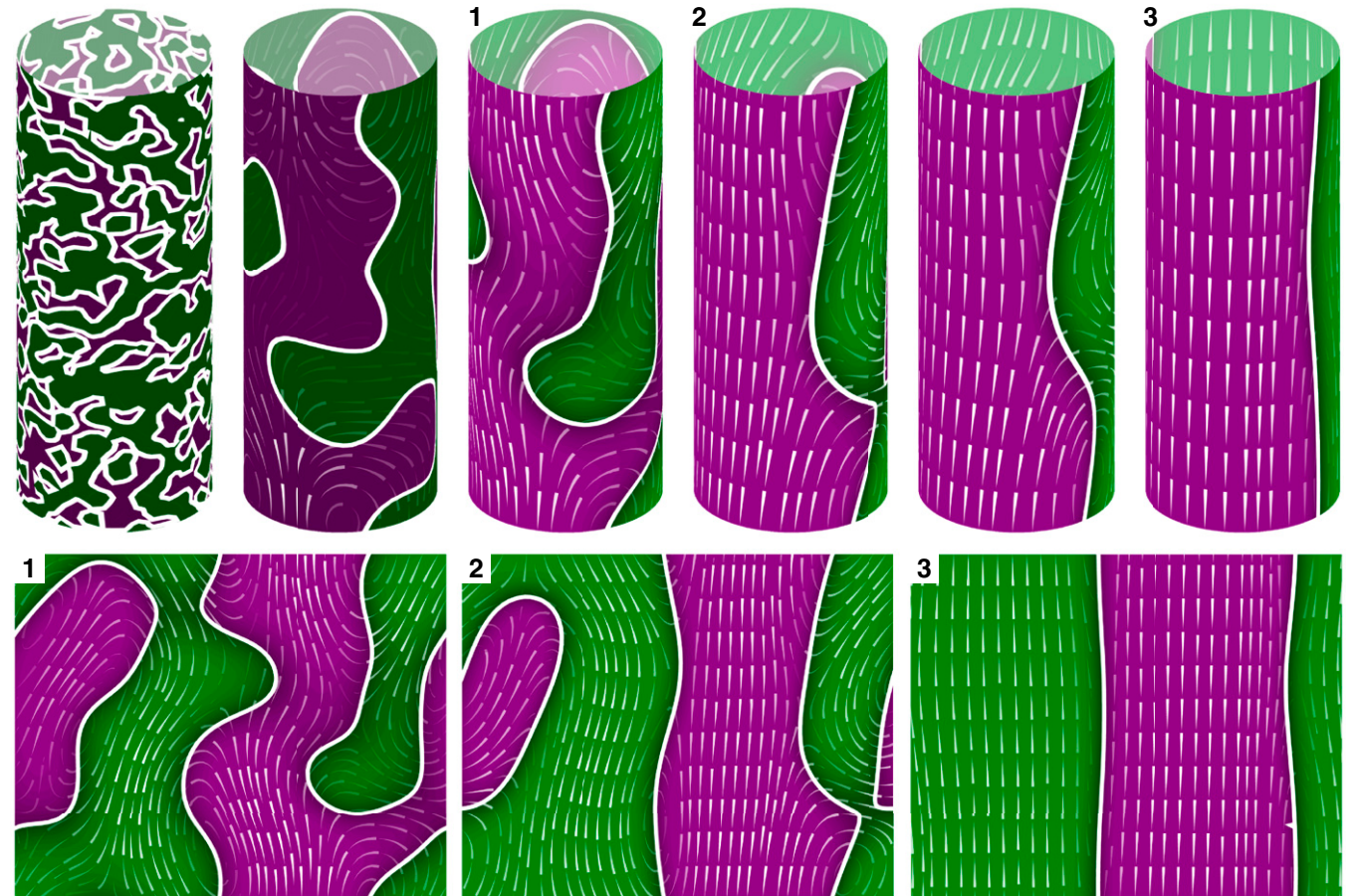
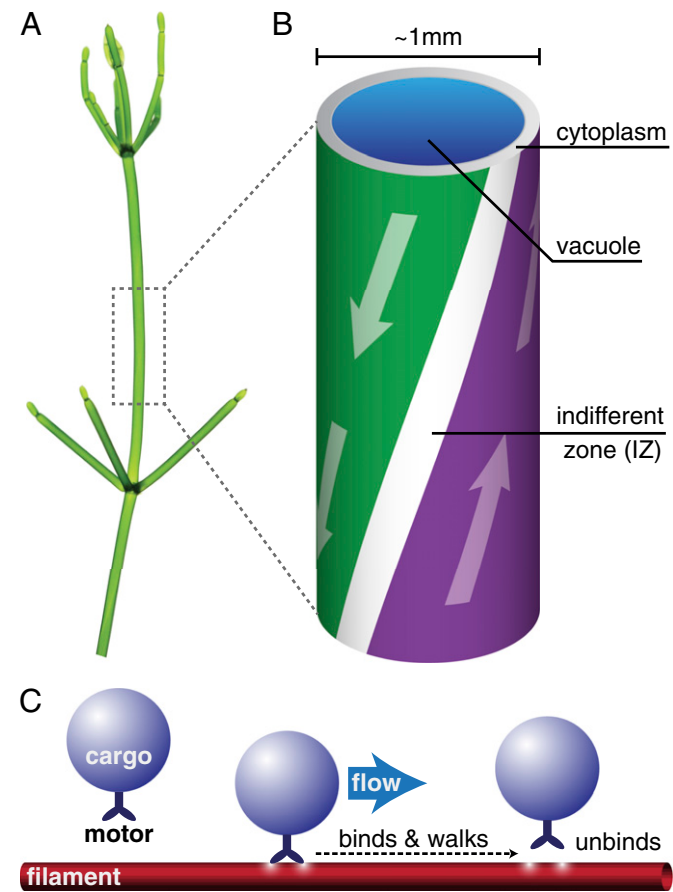
typical plant cell has between 10^3 and 10^5 plasmodesmata
 connecting it with adjacent cells
 equating to between 1 and 10 per μm

Chara fragilis



[http://www.youtube.com/watch?
feature=player_detailpage&v=kud4qUhsCyg](http://www.youtube.com/watch?feature=player_detailpage&v=kud4qUhsCyg)

Characean algae



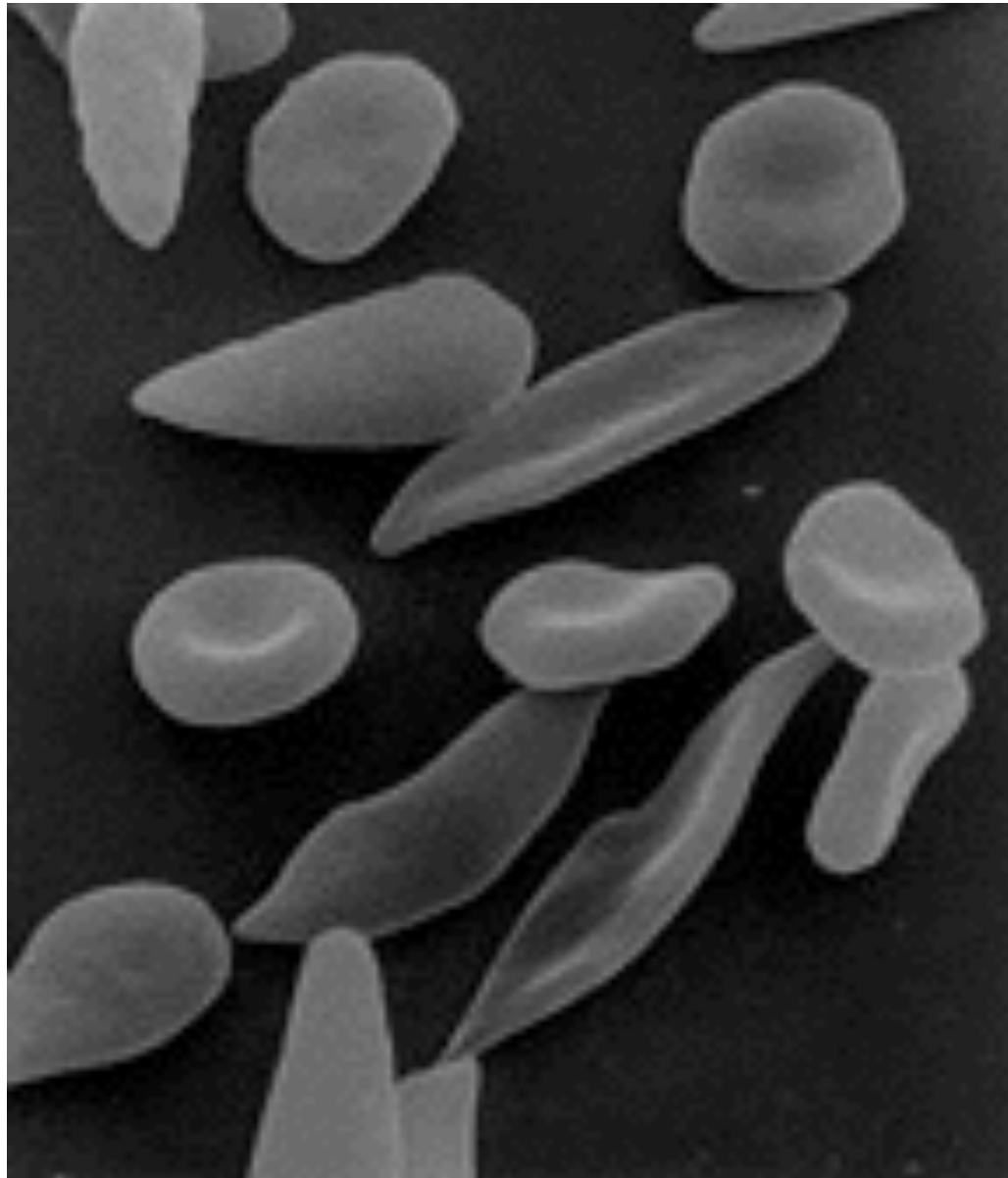
$$-\mu \nabla^2 \mathbf{u} + \mathbf{u} + \Pi_0 \mathbf{e}_z + \nabla \Pi' = |\mathbf{P}|^2 \mathbf{P},$$

$$\nabla \cdot \mathbf{u} = 0$$

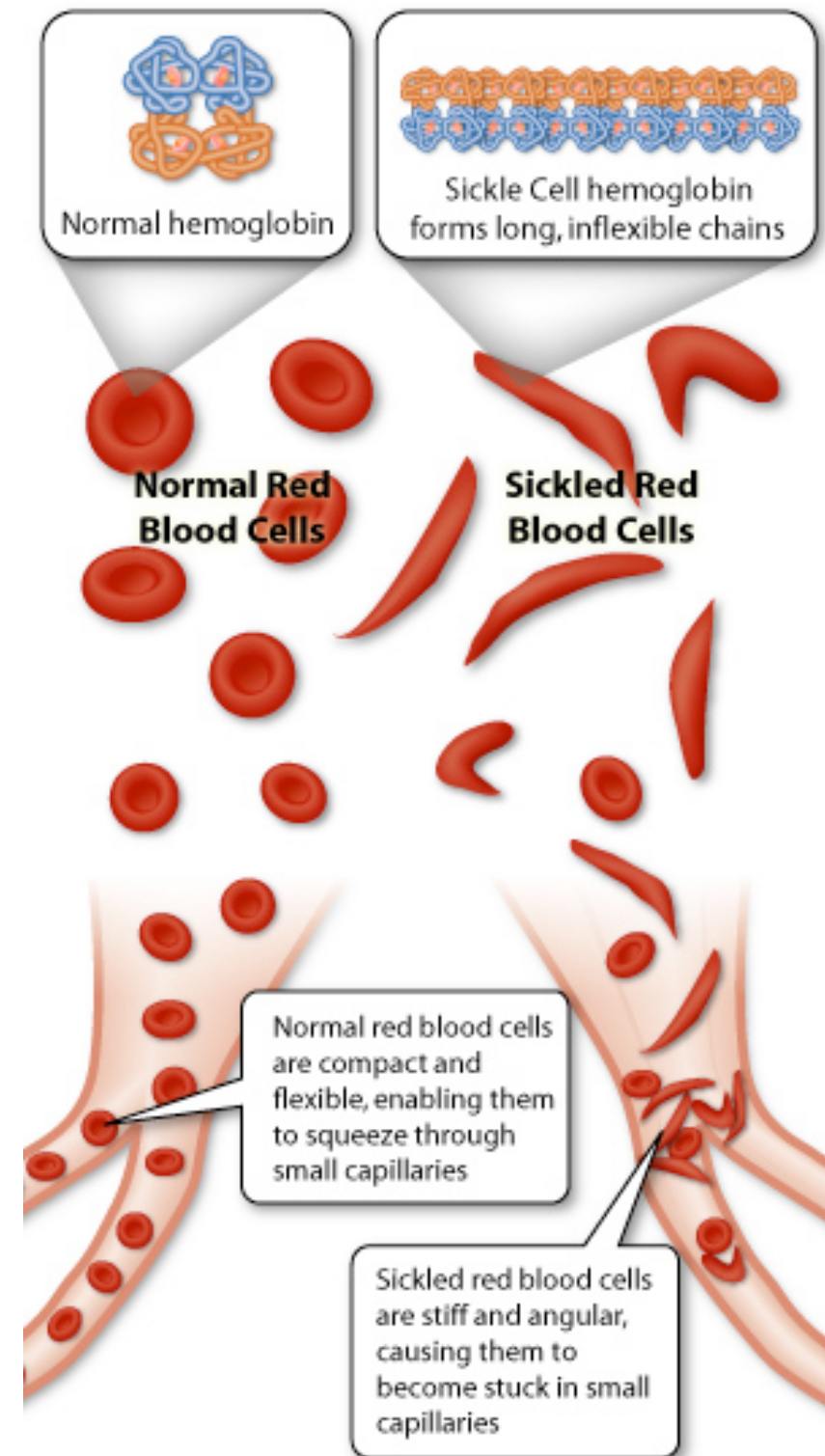
$$\frac{\partial \mathbf{P}}{\partial t} + \epsilon \mathbf{u} \cdot \nabla \mathbf{P} = d^{(s)} \nabla^2 \mathbf{P} - d^{(r)} \mathbf{P} + (\mathbb{I} - \mathbf{P}\mathbf{P}) \cdot [\epsilon (\nabla \mathbf{u}) \cdot \mathbf{P} + \alpha_p \mathbf{P} + \alpha_u \mathbf{u} - \kappa (\mathbf{P} \cdot \mathbf{d}) \mathbf{d}],$$

Blood cells: shape & function

source: wiki

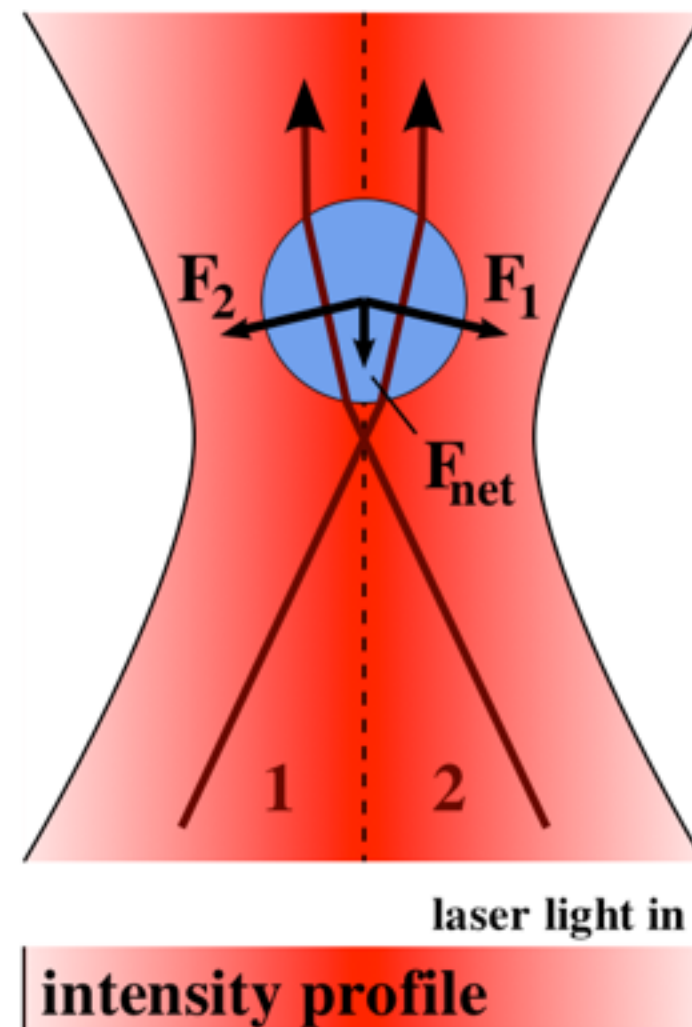
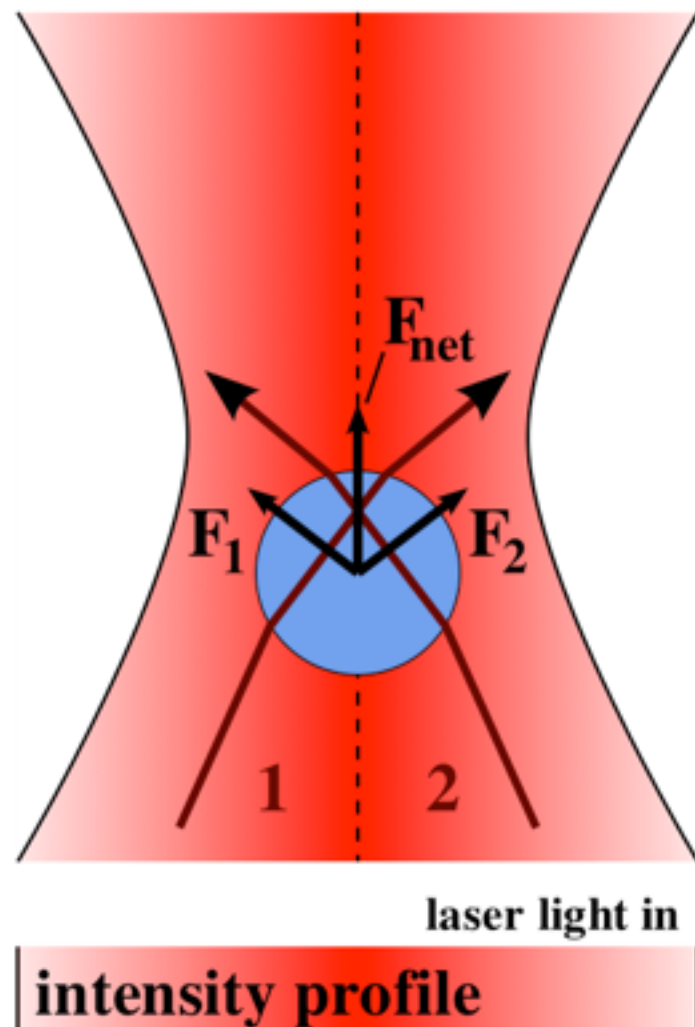


red blood cells
affected by sickle-
cell disease



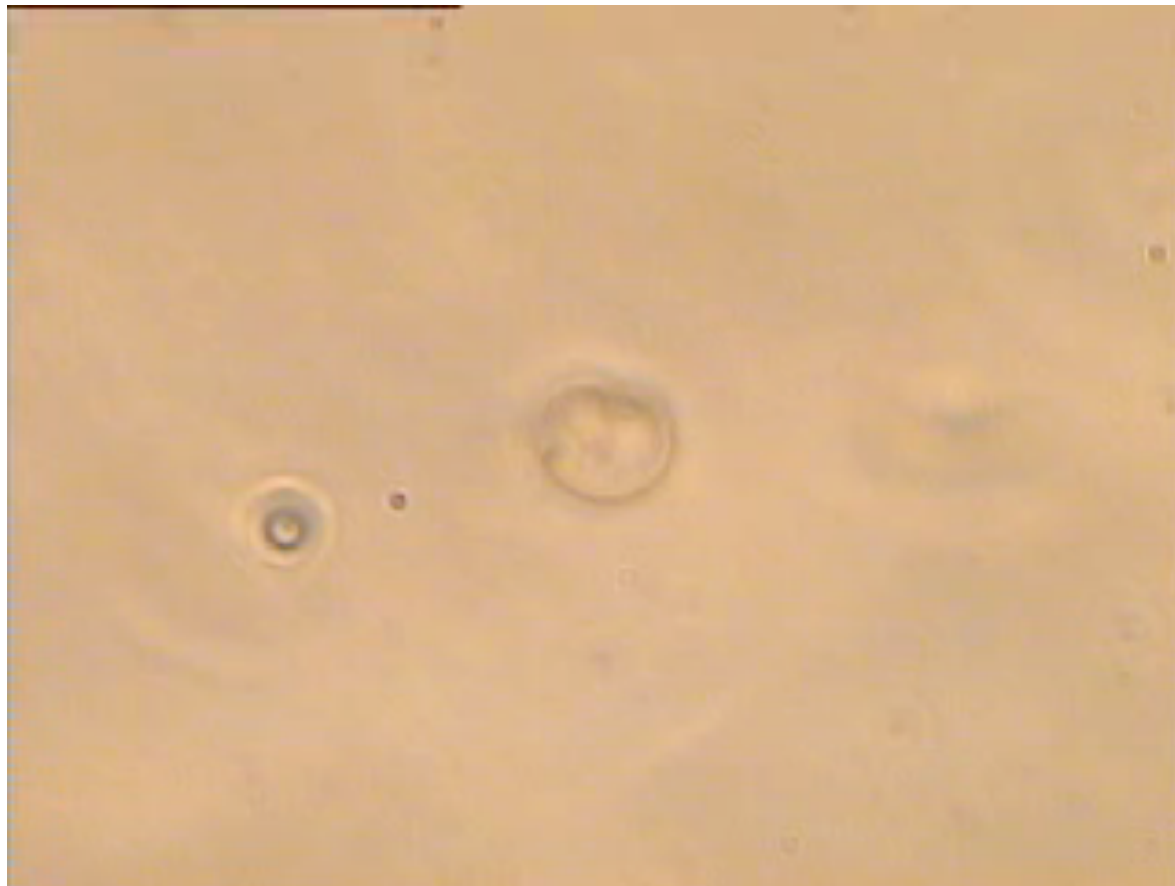
Optical tweezer

source: wiki



<http://www.nature.com/ncomms/journal/v4/n4/extref/ncomms2786-s1.swf>

Red blood cell in tweezer



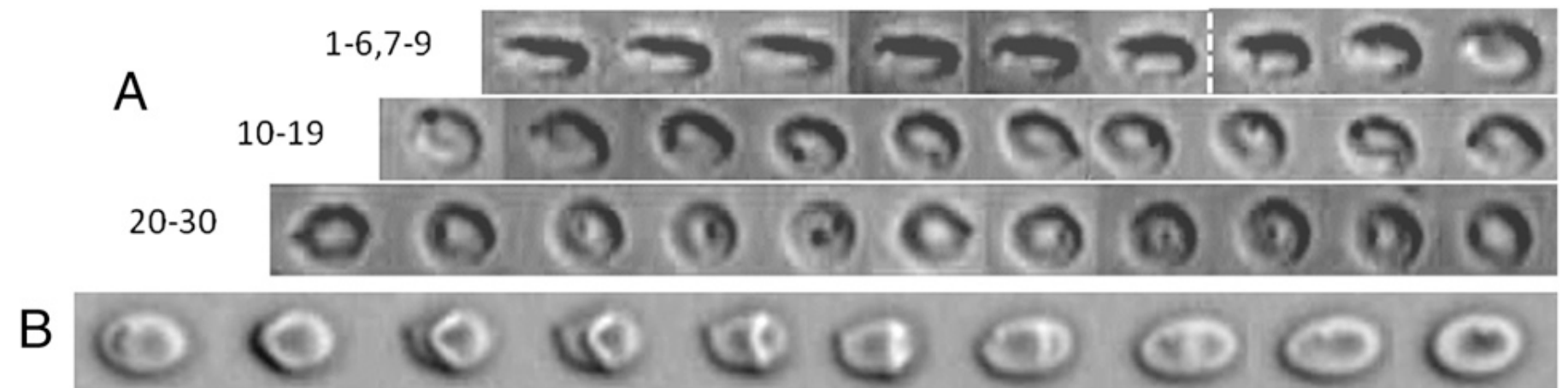
Basu et al (2011) Biophys J

dunkel@math.mit.edu

Full dynamics of a red blood cell in shear flow

Jules Dupire, Marius Socol, and Annie Viallat¹

Fig. 5. Rolling-to-tank-treading transition observed on RBCs bearing a bead; dextran $2 \cdot 10^6$ g/mol, $c = 9\%$ (wt/wt); scale bar, $8 \mu\text{m}$; top-view observation. (A) Shear rate = 3 s^{-1} . The symmetry axis of the rolling cell (images 1–7) rotates gradually (images 8–10). The spinning about the symmetry axis is detected by the bead motion (images 10–19). Finally, the streamlines change and the cell tank-treads (images 20–30). A vertical bar separates the different movements. Sequence of 46.6 s; scale bar, $7 \mu\text{m}$. (B) The tank-treading movement at the transition sometimes presents an overall rotation of part of the membrane, which behaves locally like a solid by rotating as a whole. $\dot{\gamma} = 6 \text{ s}^{-1}$, time sequence of 1.98 s.



Full dynamics of a red blood cell in shear flow

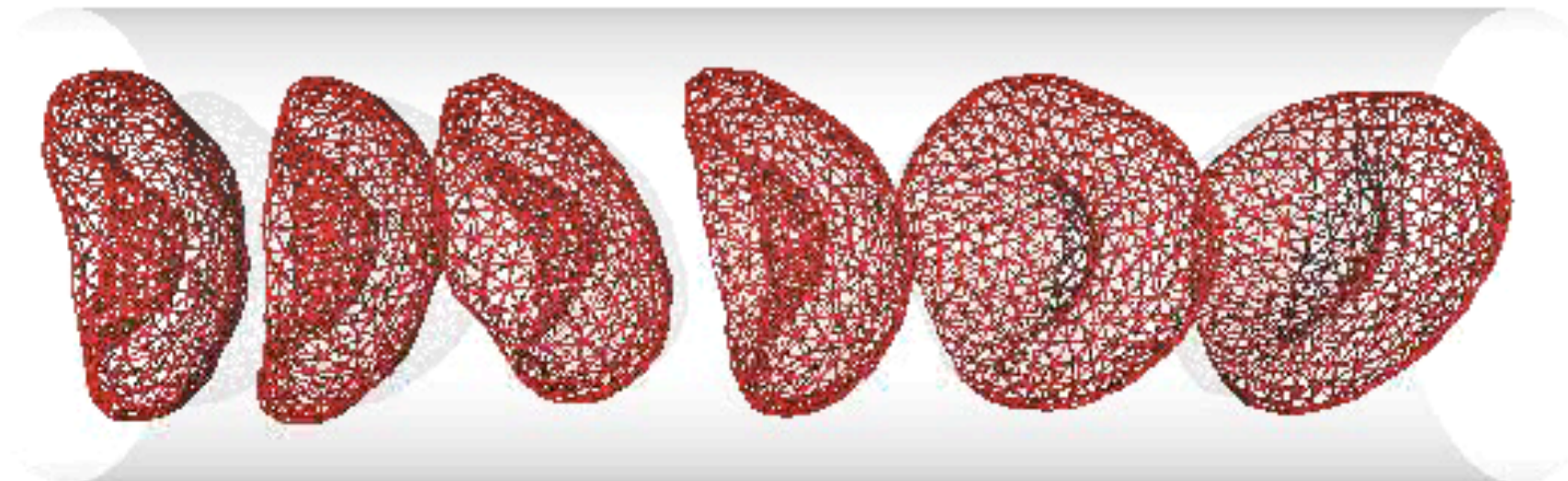
Jules Dupire, Marius Socol, and Annie Viallat¹



Rolling to Tank Treading Transition
with deformation

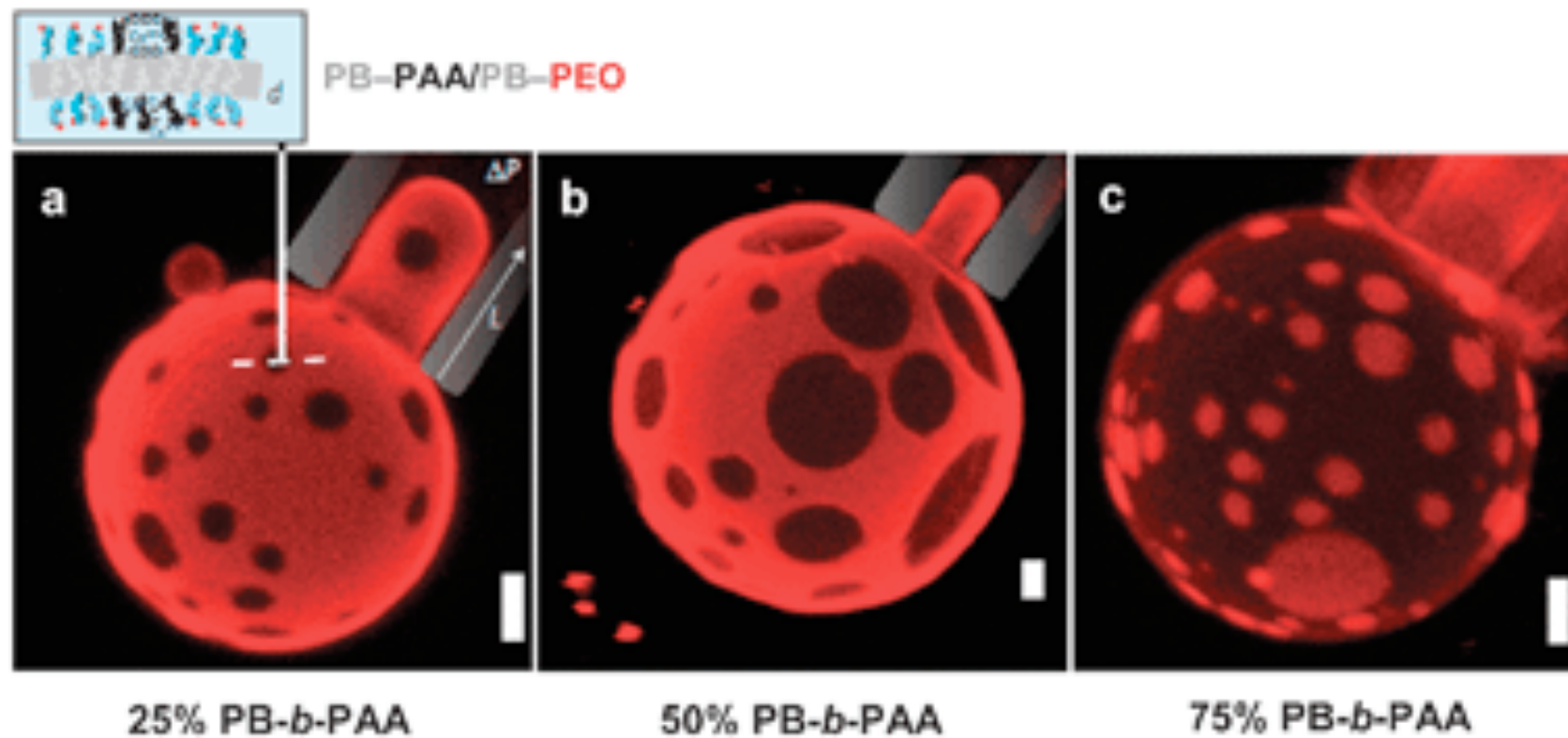
Dupire J, Socol M, Viallat A
2012

Blood cell - simulations



McWhirter et al (2012) New J Phys

Vesicles (“artificial” cells)



Mai & Eisenberg
Chem Soc Rev 2012

Membrane Viscosity Determined from Shear-Driven Flow in Giant Vesicles

Aurelia R. Honerkamp-Smith, Francis G. Woodhouse, Vasily Kantsler, and Raymond E. Goldstein

Department of Applied Mathematics and Theoretical Physics, Centre for Mathematical Sciences, University of Cambridge,
Wilberforce Road, Cambridge CB3 0WA, United Kingdom
(Received 14 January 2013; published 17 July 2013)

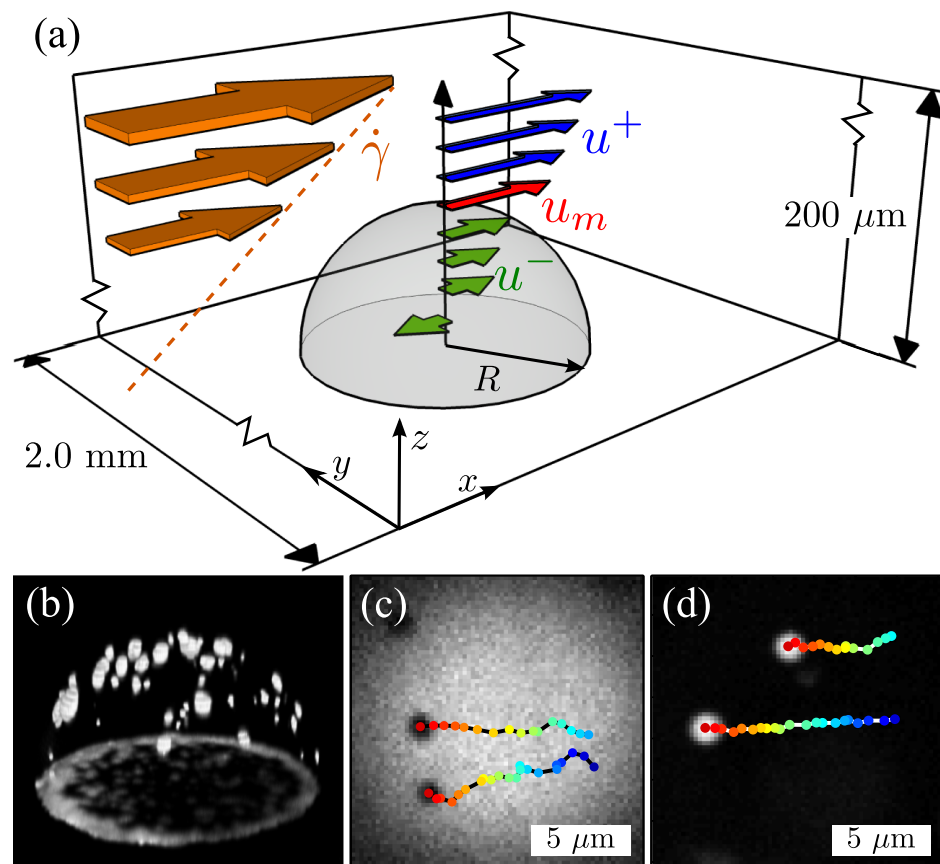


FIG. 1 (color online). Microfluidic shear experiment. (a) Schematic of the chamber (not to scale) and flows. (b) Confocal imaging reconstruction of an adhering hemispherical L_o phase vesicle with small L_d domains visible on its surface. (c)–(d) Tracking of gel domains in L_d background (c) and L_d domains in L_o background (d), flowing across the vesicle apex at $\dot{\gamma} = 2.6 \text{ s}^{-1}$ (tracks color-coded in time over $\sim 2.6 \text{ s}$).

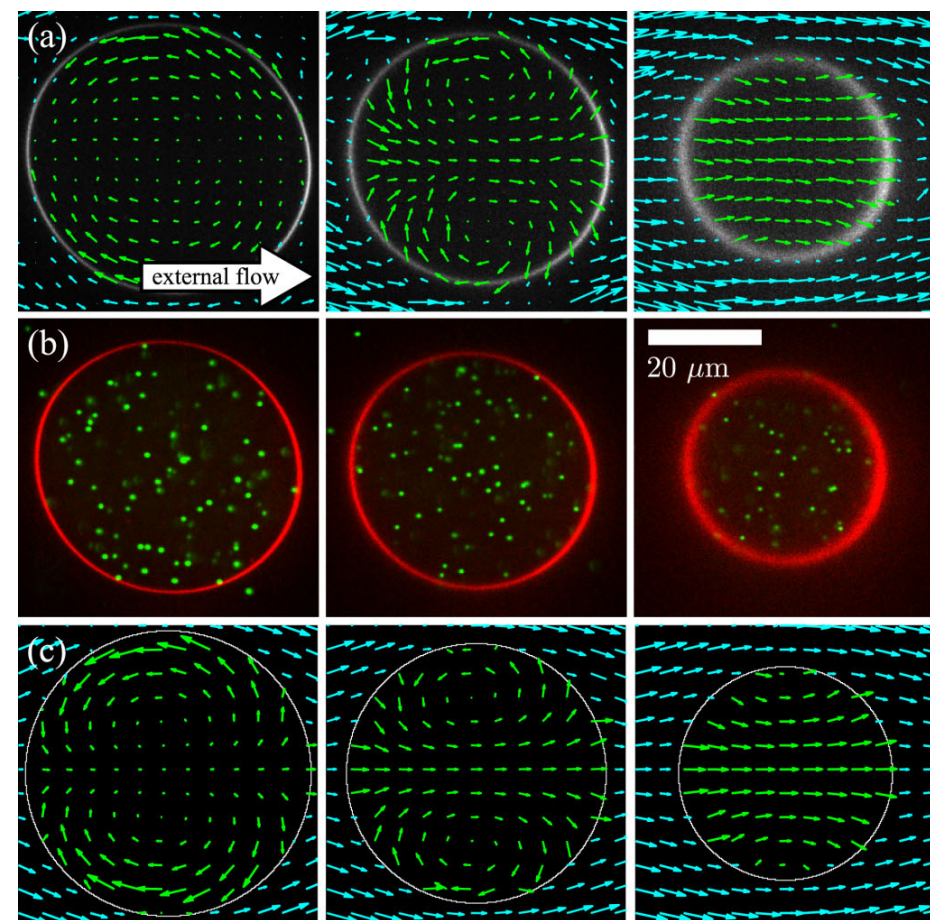


FIG. 2 (color online). Flow fields inside an adhering vesicle in shear. (a) Experimental two-dimensional PIV velocity fields at heights $z/R = 0.26, 0.47, 0.71$ above coverslip. (b) Confocal slices at same fractional heights as (a) show vesicle (red) containing fluorescent microspheres. (c) Theoretical two-dimensional velocity fields [25] for a sheared hemispherical vesicle at $z/R = 0.3, 0.5, 0.7$. Interior and exterior PIV vectors

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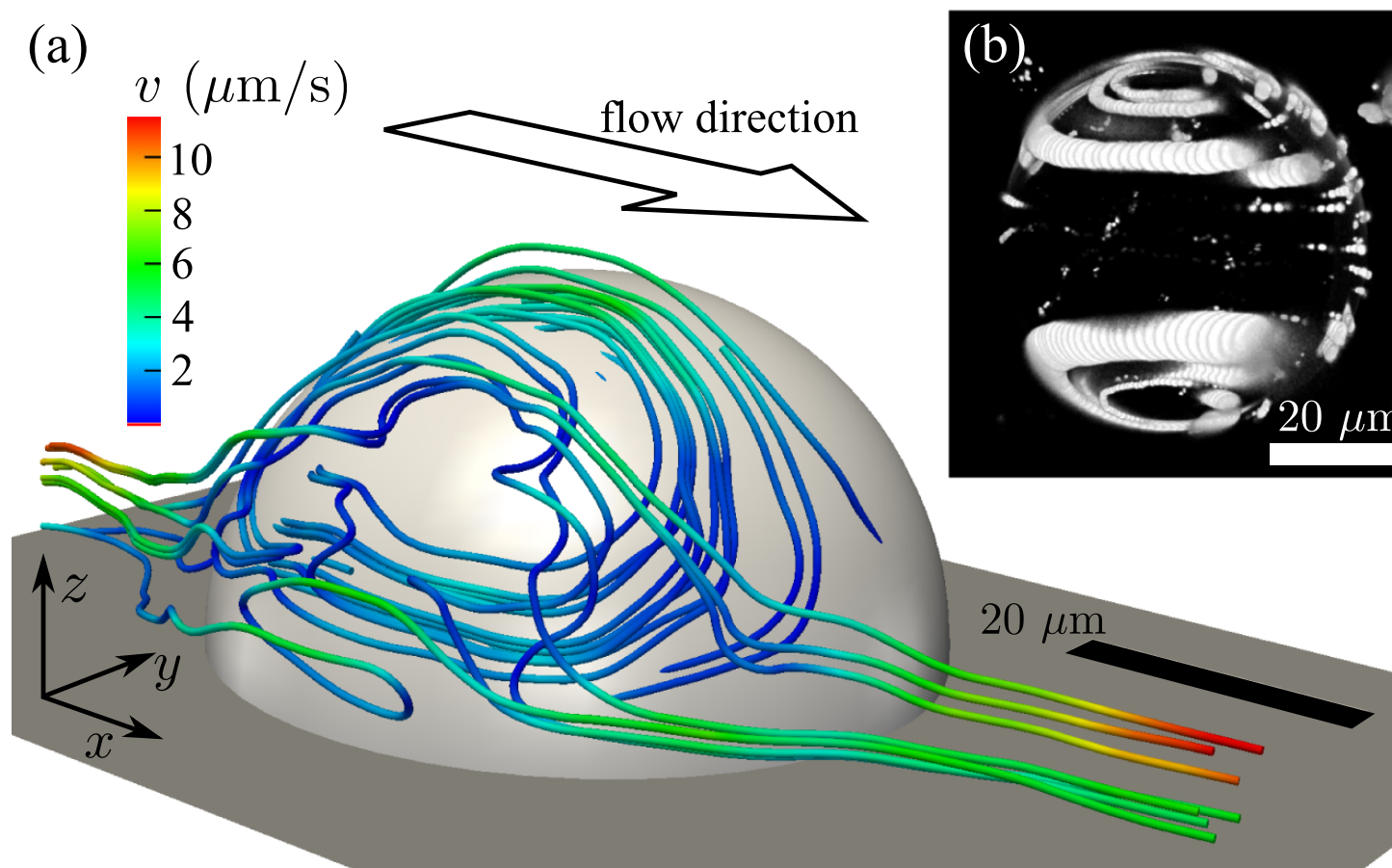


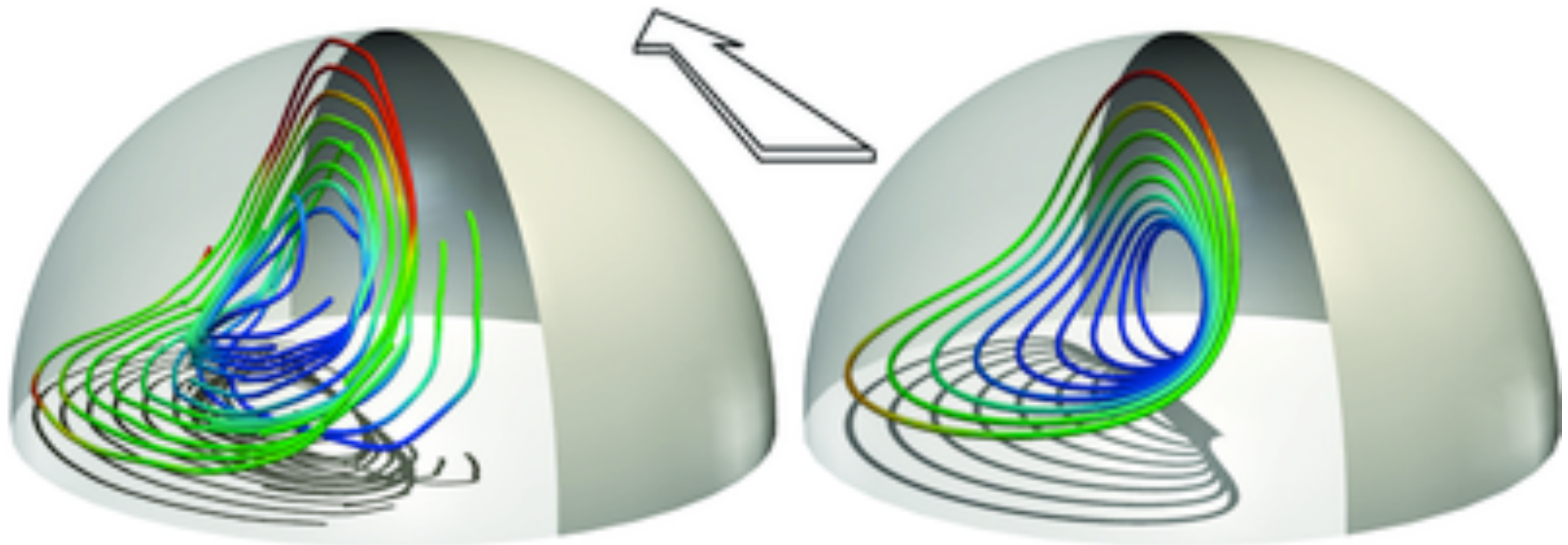
FIG. 3 (color online). Membrane and external flows. (a) Selected external streamlines along one side of an L_o vesicle in shear flow, showing closed orbits above the surface. (b) Time-lapse confocal stack of an L_o vesicle, viewed from above, illustrating circulation of L_d domains.

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Dynamics of a vesicle in general flow

J. Deschamps, V. Kantsler, E. Segre, and V. Steinberg¹

Department of Physics of Complex Systems, Weizmann Institute of Science, Rehovot, 76100 Israel

11444-11447 | PNAS | July 14, 2009 | vol. 106 | no. 28

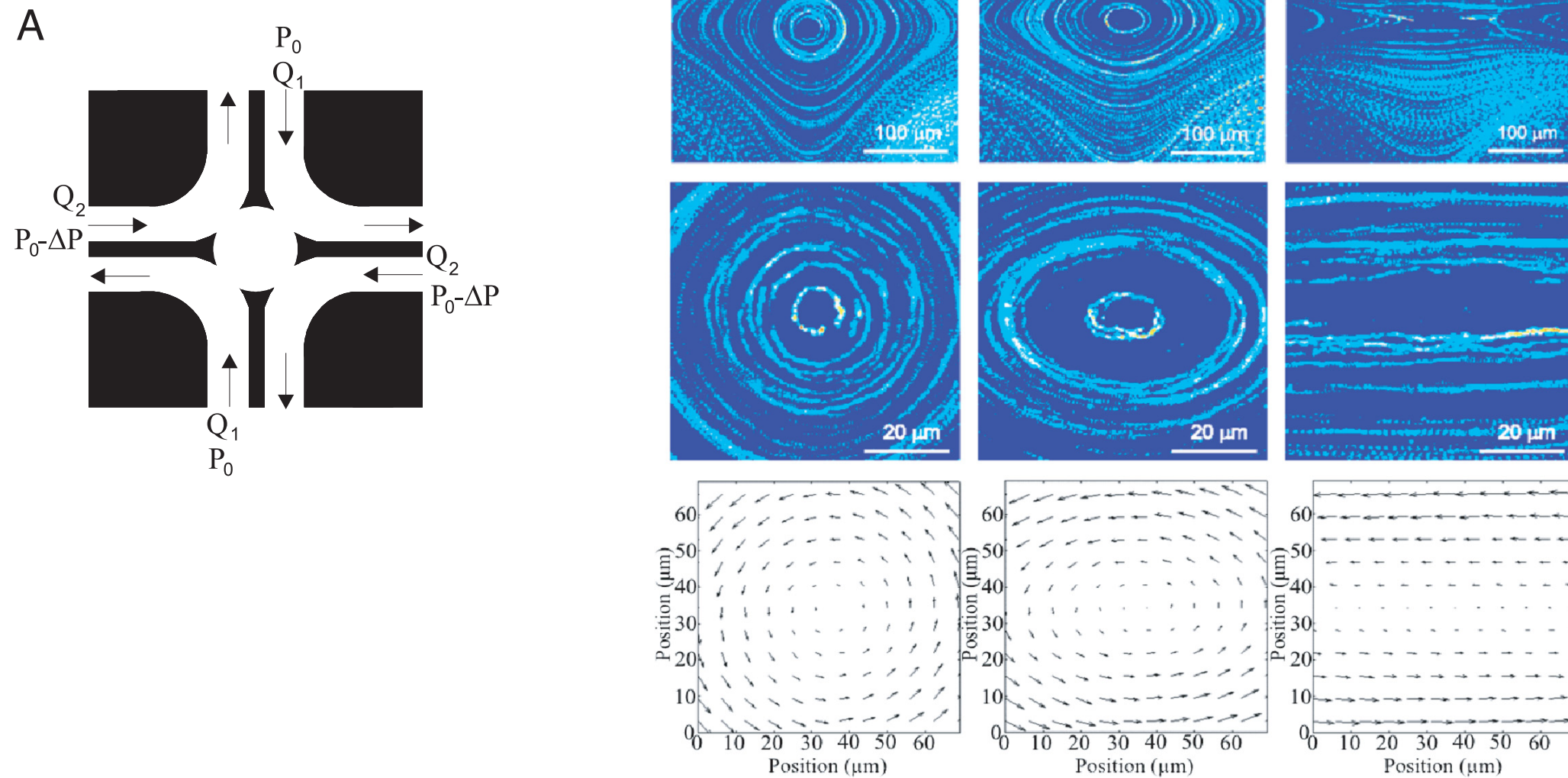


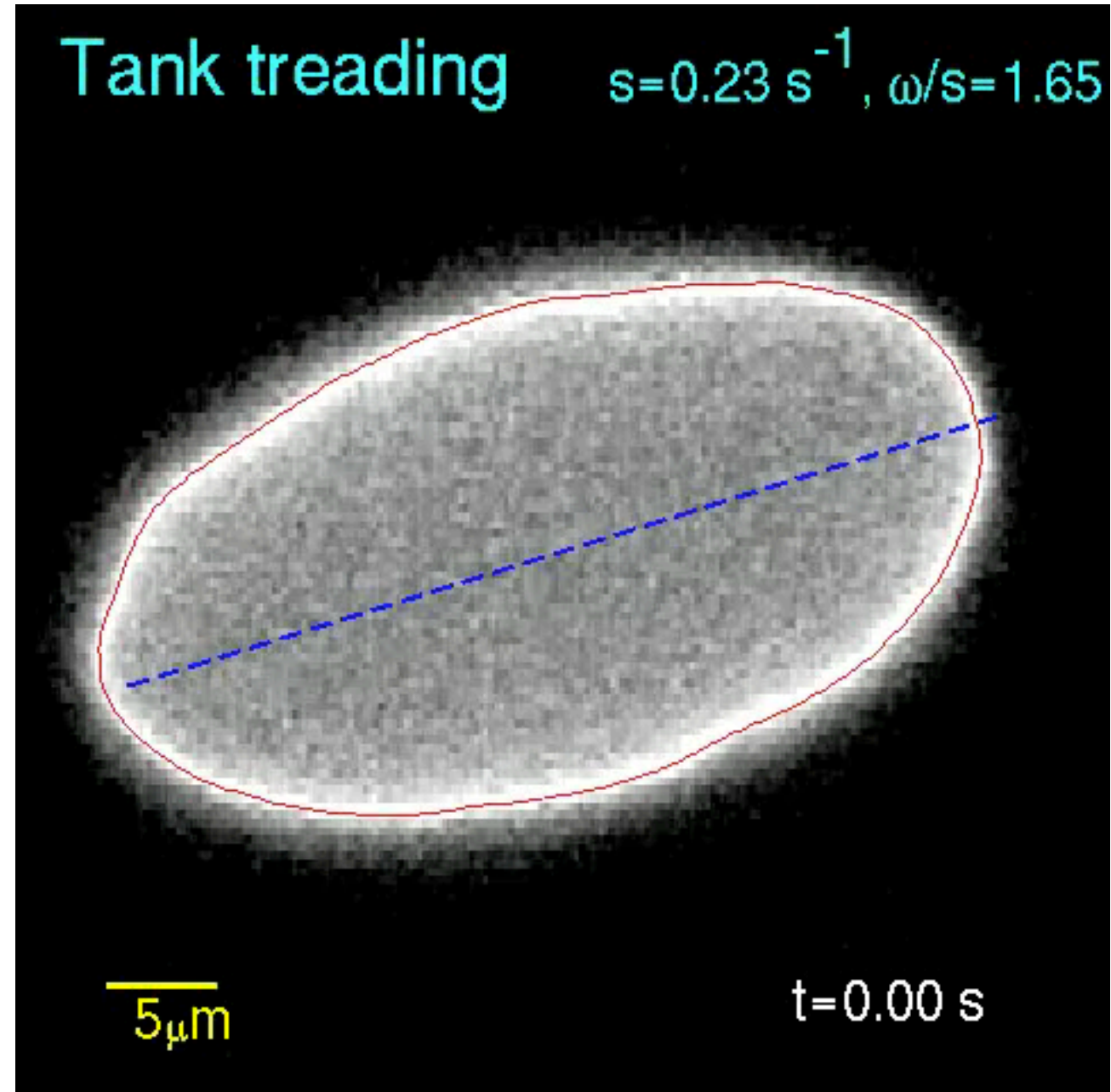
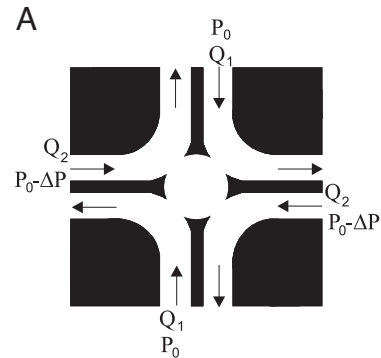
Fig. 2. (A) Experimental streamlines images of the velocity fields for pure rotational (first column, $\omega/s = 43$), mixed (second column, $\omega/s = 2.6$) and pure shear (third, $\omega/s = 1$) flows; (B) Zoom of the same experimental flows; (C) velocity vector field representation of the same flows (PTV).

Dynamics of a vesicle in general flow

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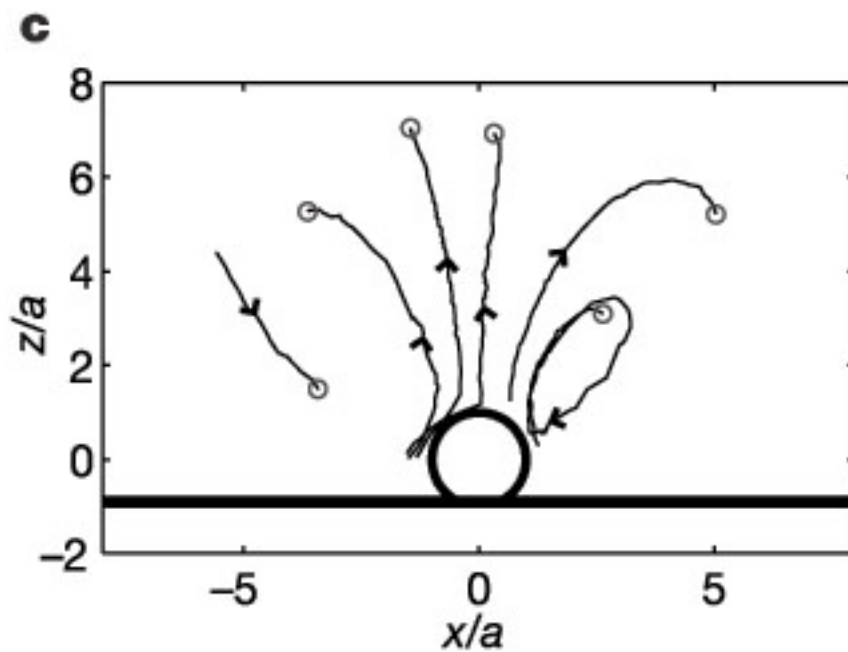
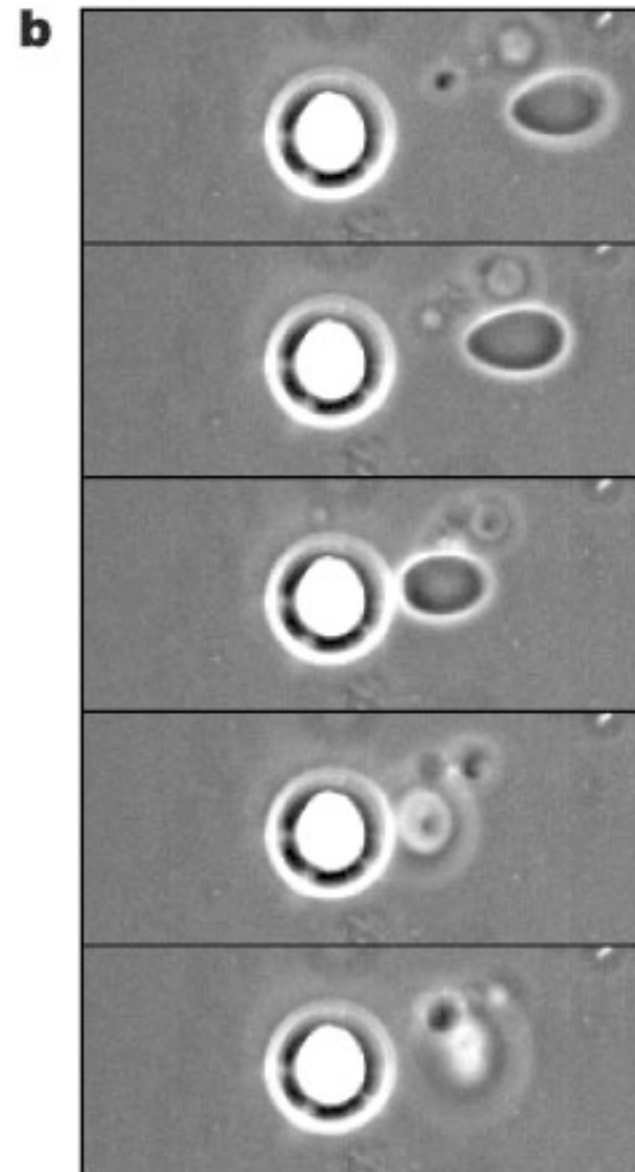
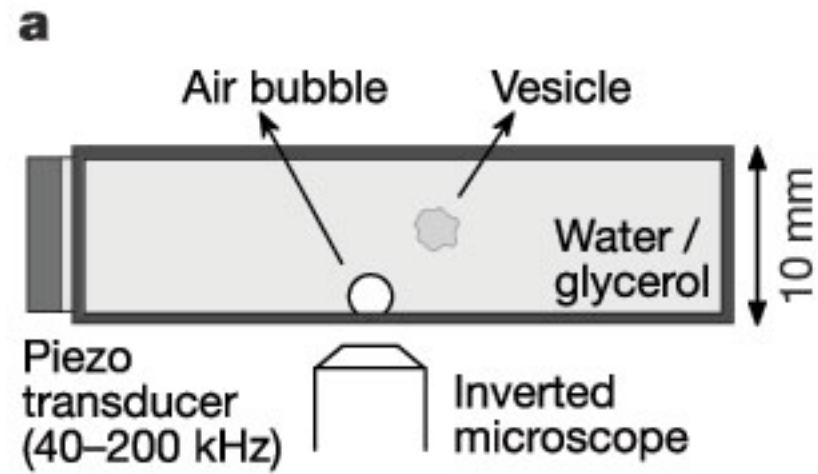
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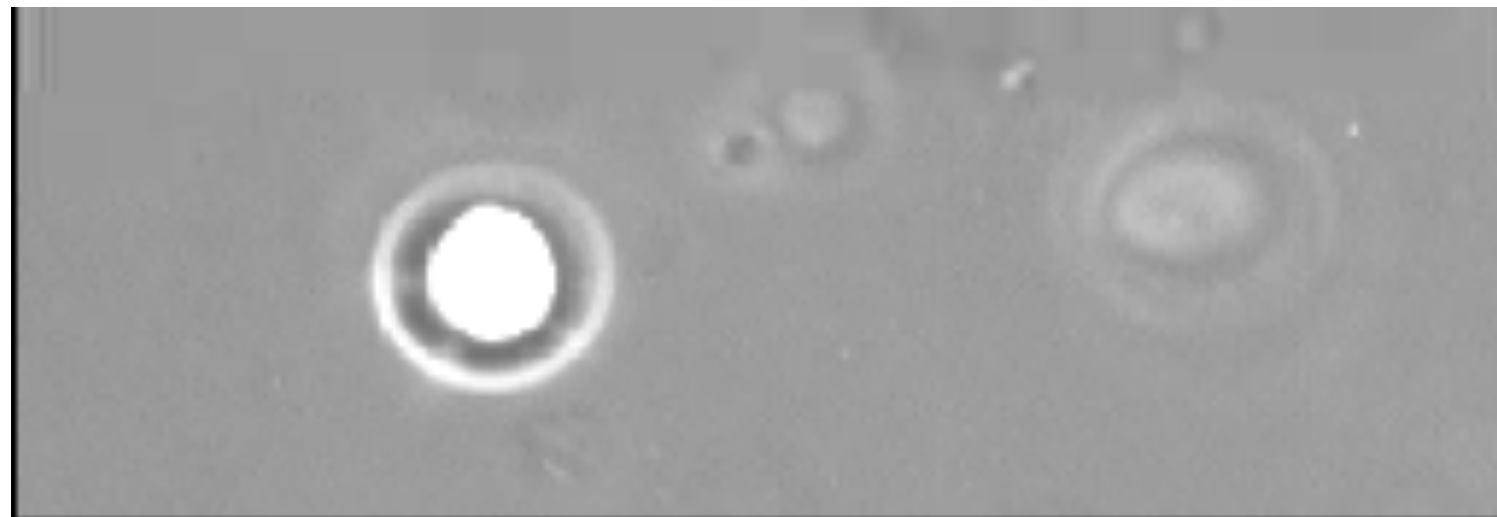


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Controlled vesicle deformation and lysis by single oscillating bubbles

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Faculty of Applied Physics, University of Twente, PO Box 217, 7500AE Enschede, The Netherlands





NATURE | VOL 423 | 8 MAY 2003 | www.nature.com/nature

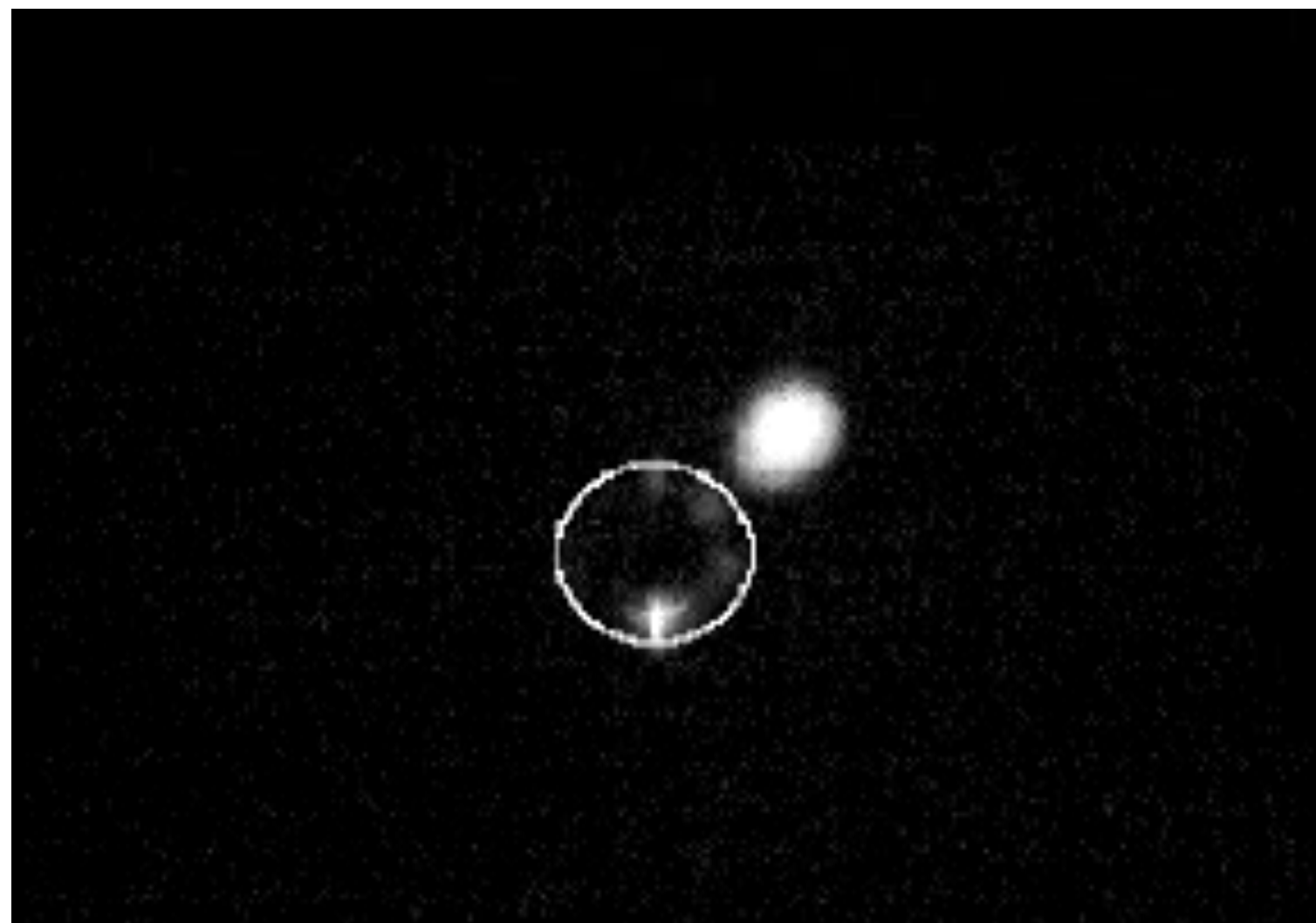
letters to nature

.....

Controlled vesicle deformation and lysis by single oscillating bubbles

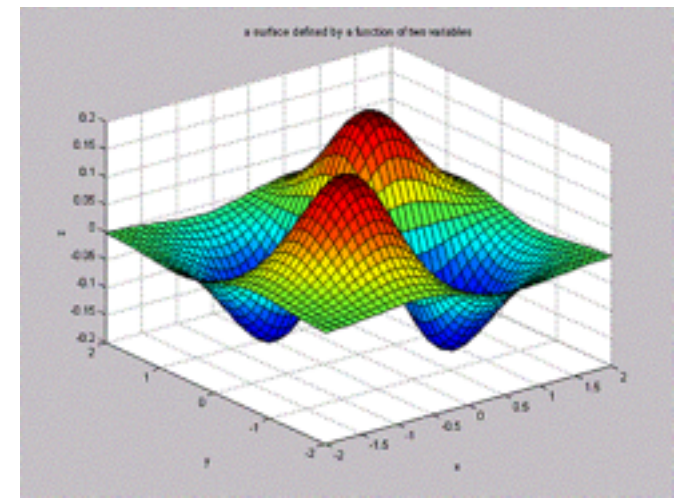
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Membranes



The discussion in this section builds on the review article [Sei97] and the textbook [OLXY99].

Reminder: 2D differential geometry

We consider an orientable surface in \mathbb{R}^3 . Possible local parameterizations are

$$\mathbf{F}(s_1, s_2) \in \mathbb{R}^3 \quad (3.1)$$

where $(s_1, s_2) \in U \subseteq \mathbb{R}^2$. Alternatively, if one chooses Cartesian coordinates $(s_1, s_2) = (x, y)$, then it suffices to specify

$$z = f(x, y) \quad (3.2a)$$

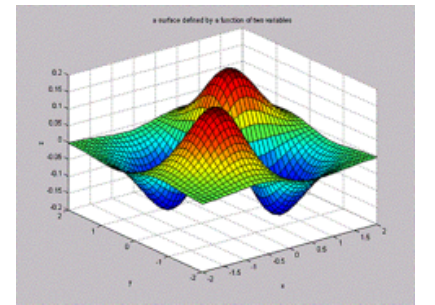
or, equivalently, the implicit representation

$$\Phi(x, y, z) = z - f(x, y). \quad (3.2b)$$

The vector representation (3.1) can be related to the ‘height’ representation (3.2a) by

$$\mathbf{F}(x, y) = \begin{pmatrix} x \\ y \\ f(x, y) \end{pmatrix} \quad (3.3)$$

Surface metric tensor



$$\mathbf{F}(x, y) = \begin{pmatrix} x \\ y \\ f(x, y) \end{pmatrix} \quad (3.3)$$

Denoting derivatives by $\mathbf{F}_i = \partial_{s_i} \mathbf{F}$, we introduce the surface metric tensor $g = (g_{ij})$ by

$$g_{ij} = \mathbf{F}_i \cdot \mathbf{F}_j, \quad (3.4a)$$

abbreviate its determinant by

$$|g| := \det g, \quad (3.4b)$$

and define the associated Laplace-Beltrami operator ∇^2 by

$$\nabla^2 h = \frac{1}{\sqrt{|g|}} \partial_i (g_{ij}^{-1} \sqrt{|g|} \partial_j h), \quad (3.4c)$$

for some function $h(s_1, s_2)$.

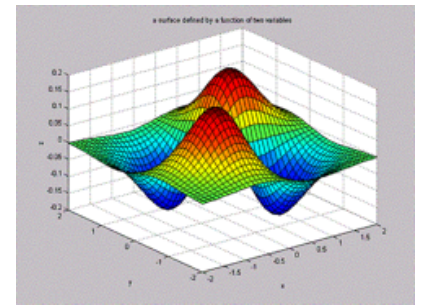
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abbreviate its determinant by

$$|g| := \det g, \quad (3.4b)$$



$$\mathbf{F}_x(x, y) = \begin{pmatrix} 1 \\ 0 \\ f_x \end{pmatrix}, \quad \mathbf{F}_y(x, y) = \begin{pmatrix} 0 \\ 1 \\ f_y \end{pmatrix} \quad (3.5)$$

and, hence, the metric tensor

$$g = (g_{ij}) = \begin{pmatrix} \mathbf{F}_x \cdot \mathbf{F}_x & \mathbf{F}_x \cdot \mathbf{F}_y \\ \mathbf{F}_y \cdot \mathbf{F}_x & \mathbf{F}_y \cdot \mathbf{F}_y \end{pmatrix} = \begin{pmatrix} 1 + f_x^2 & f_x f_y \\ f_y f_x & 1 + f_y^2 \end{pmatrix} \quad (3.6a)$$

and its determinant

$$|g| = 1 + f_x^2 + f_y^2, \quad (3.6b)$$

where $f_x = \partial_x f$ and $f_y = \partial_y f$. For later use, we still note that the inverse of the metric tensor is given by

$$g^{-1} = (g_{ij}^{-1}) = \frac{1}{1 + f_x^2 + f_y^2} \begin{pmatrix} 1 + f_y^2 & -f_x f_y \\ -f_y f_x & 1 + f_x^2 \end{pmatrix}. \quad (3.6c)$$

Surface normal & curvature

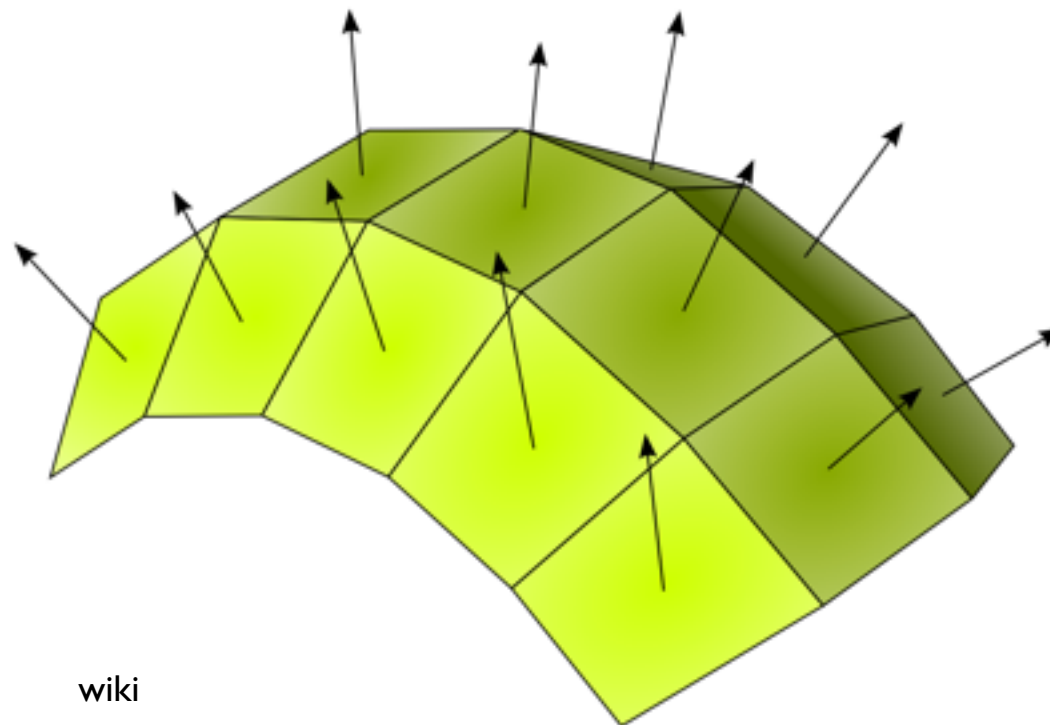
Assuming the surface is regular at (s_1, s_2) , which just means that the tangent vectors \mathbf{F}_1 and \mathbf{F}_2 are linearly independent, the local unit normal vector is defined by

$$\mathbf{N} = \frac{\mathbf{F}_1 \wedge \mathbf{F}_2}{\|\mathbf{F}_1 \wedge \mathbf{F}_2\|}. \quad (3.7)$$

In terms of the Cartesian parameterization, this can also be rewritten as

$$\mathbf{N} = \frac{\nabla\Phi}{\|\nabla\Phi\|} = \frac{1}{\sqrt{1 + f_x^2 + f_y^2}} \begin{pmatrix} -f_x \\ -f_y \\ 1 \end{pmatrix}. \quad (3.8)$$

Here, we have adopted the convention that $\{\mathbf{F}_1, \mathbf{F}_2, \mathbf{N}\}$ form a right-handed system.



Surface normal & curvature

Assuming the surface is regular at (s_1, s_2) , which just means that the tangent vectors \mathbf{F}_1 and \mathbf{F}_2 are linearly independent, the local unit normal vector is defined by

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Here, we have adopted the convention that $\{\mathbf{F}_1, \mathbf{F}_2, \mathbf{N}\}$ form a right-handed system.

To formulate ‘geometric’ energy functionals for membranes, we still require the concept of curvature, which quantifies the local bending of the membrane. We define a 2×2 -curvature tensor $R = (R_{ij})$ by

$$R_{ij} = \mathbf{N} \cdot (\mathbf{F}_{ij}) \quad (3.9)$$

and local *mean curvature* H and local *Gauss curvature* K by

$$H = \frac{1}{2} \text{tr}(g^{-1} \cdot R), \quad K = \det(g^{-1} \cdot R). \quad (3.10)$$

Adopting the Cartesian representation (3.2a), we have

$$\mathbf{F}_{xx} = \begin{pmatrix} 0 \\ 0 \\ f_{xx} \end{pmatrix}, \quad \mathbf{F}_{xy} = \mathbf{F}_{yx} = \begin{pmatrix} 0 \\ 0 \\ f_{xy} \end{pmatrix}, \quad \mathbf{F}_{yy} = \begin{pmatrix} 0 \\ 0 \\ f_{yy} \end{pmatrix} \quad (3.11a)$$

Surface normal & curvature

yielding the curvature tensor

$$(R_{ij}) = \begin{pmatrix} \mathbf{N} \cdot \mathbf{F}_{xx} & \mathbf{N} \cdot \mathbf{F}_{xy} \\ \mathbf{N} \cdot \mathbf{F}_{yx} & \mathbf{N} \cdot \mathbf{F}_{yy} \end{pmatrix} = \frac{1}{\sqrt{1 + f_x^2 + f_y^2}} \begin{pmatrix} f_{xx} & f_{xy} \\ f_{yx} & f_{yy} \end{pmatrix} \quad (3.11b)$$

Denoting the eigenvalues of the matrix $g^{-1} \cdot R$ by κ_1 and κ_2 , we obtain for the mean curvature

$$H = \frac{1}{2} (\kappa_1 + \kappa_2) = \frac{(1 + f_y^2)f_{xx} - 2f_x f_y f_{xy} + (1 + f_x^2)f_{yy}}{2(1 + f_x^2 + f_y^2)^{3/2}} \quad (3.12)$$

and for the Gauss curvature

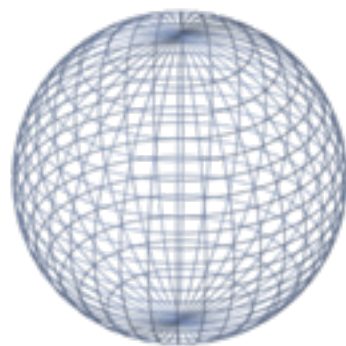
$$K = \kappa_1 \cdot \kappa_2 = \frac{f_{xx}f_{yy} - f_{xy}^2}{(1 + f_x^2 + f_y^2)^2}. \quad (3.13)$$

Gauss-Bonnet Theorem

$$\int_M K dA + \oint_{\partial M} k_g ds = 2\pi \chi(M). \quad (3.14)$$

Gauss curvature geodesic curvature Euler characteristic

$\chi(M) = 2 - 2g$, where g is the *genus*



2



0






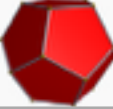

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Gauss-Bonnet Theorem

$$\int_M K \, dA + \oint_{\partial M} k_g \, ds = 2\pi \chi(M). \quad (3.14)$$

Convex polyhedra

Name	Image	Vertices V	Edges E	Faces F	Euler characteristic: $V - E + F$
Tetrahedron		4	6	4	2
Hexahedron or cube		8	12	6	2
Octahedron		6	12	8	2
Dodecahedron		20	30	12	2
Icosahedron		12	30	20	2

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Minimal surfaces

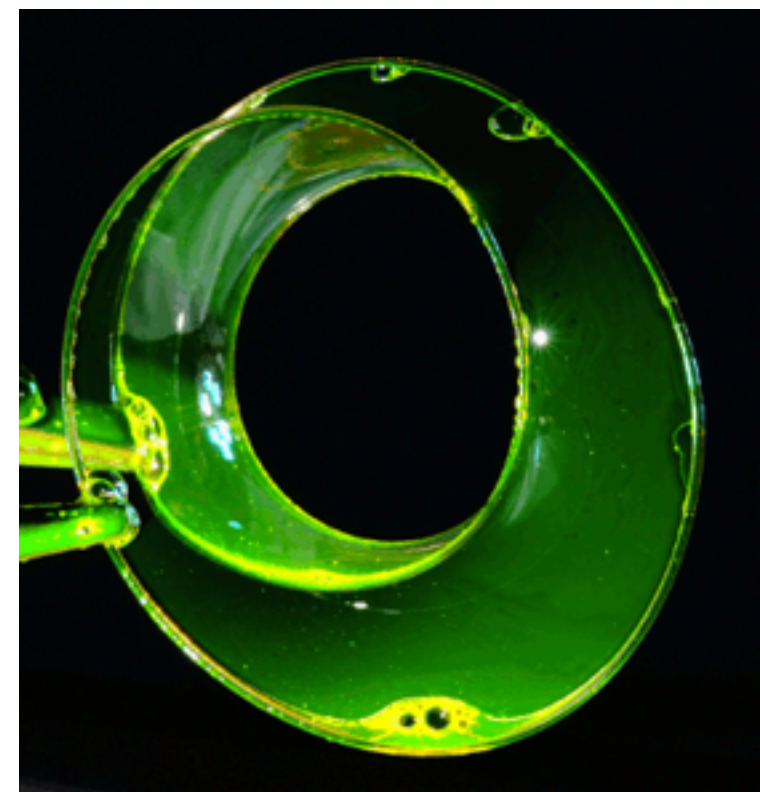
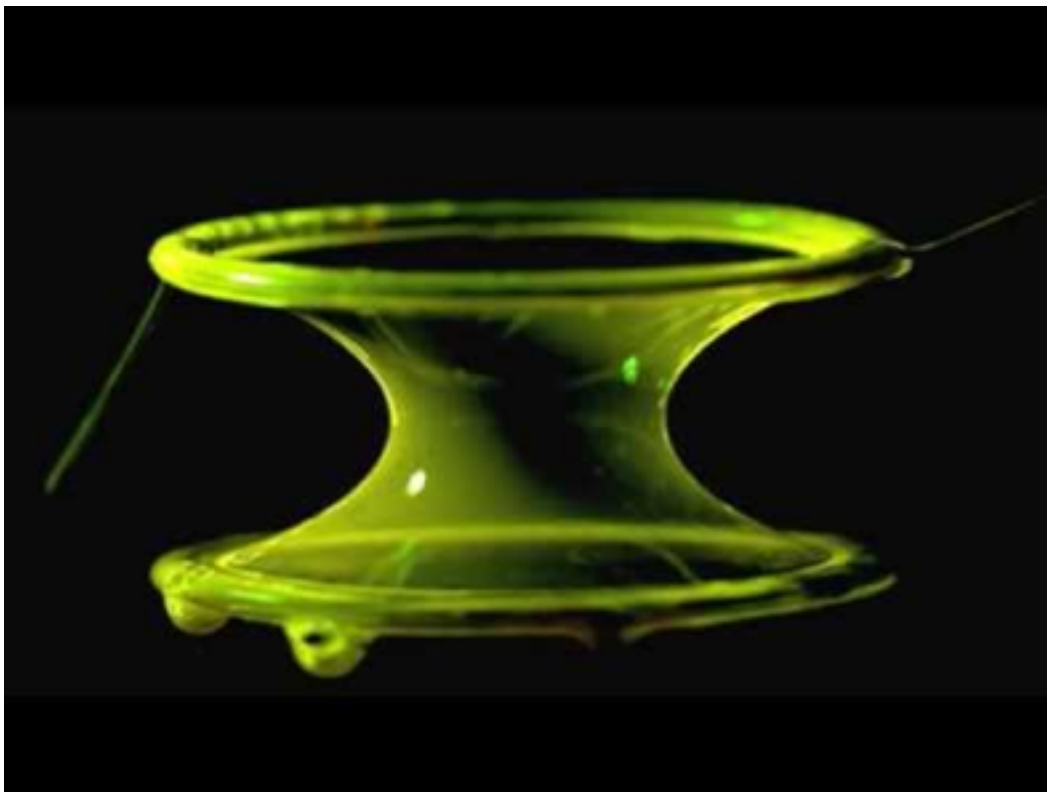
Minimal surfaces are surfaces that minimize the area within a given contour ∂M ,

$$A(M|\partial M) = \int_M dA = \min! \quad (3.15)$$

Assuming a Cartesian parameterization $z = f(x, y)$ and abbreviating $f_i = \partial_i f$ as before, we have

$$dA = \sqrt{|g|} dx dy = \sqrt{1 + f_x^2 + f_y^2} dx dy =: \mathcal{L} dx dy, \quad (3.16)$$

catenoid



Goldstein lab (Cambridge)

Minimal surfaces

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$$dA = \sqrt{|g|} dx dy = \sqrt{1 + f_x^2 + f_y^2} dx dy =: \mathcal{L} dx dy, \quad (3.16)$$

and the minimum condition (3.15) can be expressed in terms of the Euler-Lagrange equations

$$0 = \frac{\delta A}{\delta f} = -\partial_i \frac{\partial \mathcal{L}}{\partial f_i}. \quad (3.17)$$

Inserting the Lagrangian $\mathcal{L} = \sqrt{|g|}$, one finds

$$0 = - \left[\partial_x \left(\frac{f_x}{\sqrt{1 + f_x^2 + f_y^2}} \right) + \partial_y \left(\frac{f_y}{\sqrt{1 + f_x^2 + f_y^2}} \right) \right] \quad (3.18)$$

which may be recast in the form

$$0 = \frac{(1 + f_y^2)f_{xx} - 2f_x f_y f_{xy} + (1 + f_x^2)f_{yy}}{(1 + f_x^2 + f_y^2)^{3/2}} = -2H. \quad (3.19)$$

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Thus, minimal surfaces satisfy

$$H = 0 \quad \Leftrightarrow \quad \kappa_1 = -\kappa_2, \quad (3.20)$$

implying that each point of a minimal surface is a saddle point.

Helfrich's model

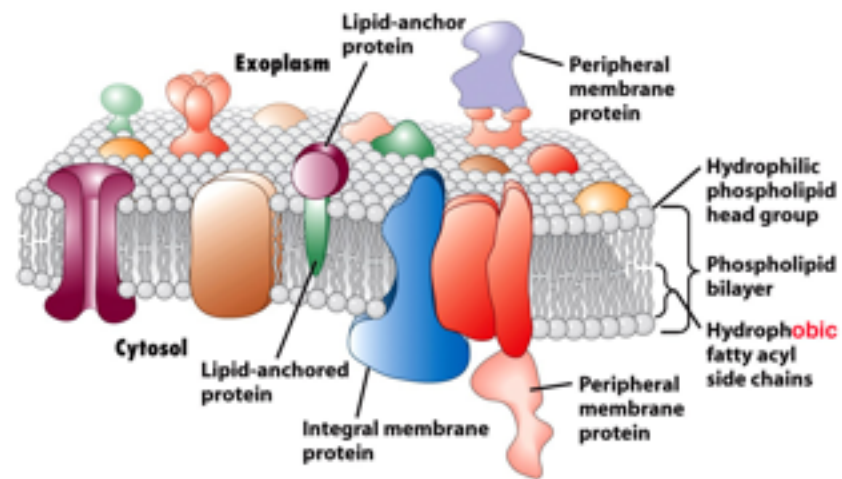
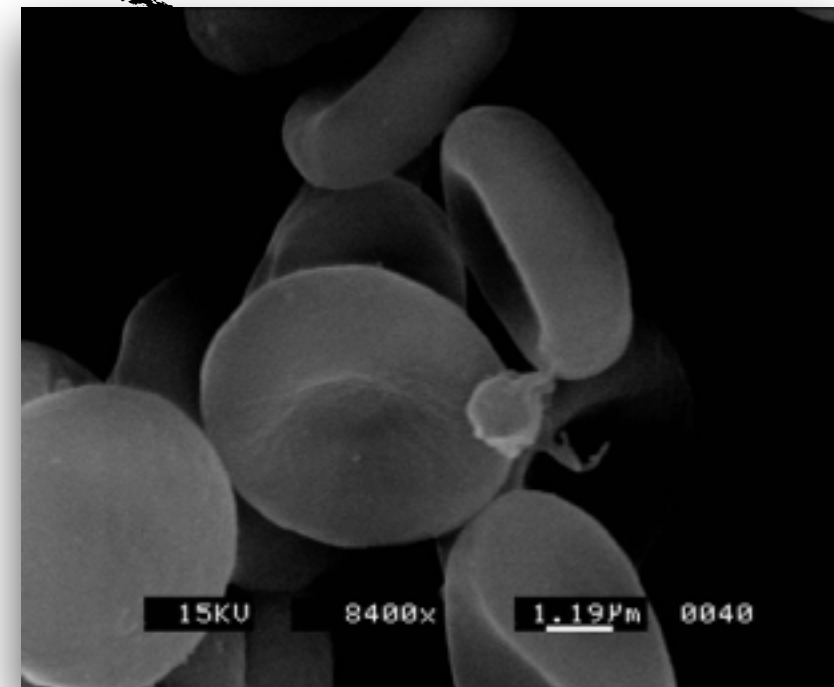


Figure 10-1
Molecular Cell Biology, Sixth Edition
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wiki



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Helfrich's model

Assuming that lipid bilayer membranes can be viewed as two-dimensional surfaces, Helfrich [Hel73] proposed in 1973 the following geometric curvature energy per unit area for a closed membrane

$$f_c = \frac{k_c}{2}(2H - c_0)^2 + k_G K, \quad (3.31)$$

where constants k_c , k_G are bending rigidities and c_0 is the spontaneous curvature of the membrane. The full free energy for a closed membrane can then be written as

$$F_c = \int dA f_c + \sigma \int dA + \Delta p \int dV, \quad (3.32)$$

where σ is the surface tension and Δp the osmotic pressure (outer pressure minus inner pressure). Minimizing F with respect to the surface shape, one finds after some heroic manipulations the shape equation²

$$\Delta p - 2\sigma H + k_c(2H - c_0)(2H^2 + c_0H - 2K) + k_c \nabla^2(2H - c_0) = 0, \quad (3.33)$$

where ∇^2 is the Laplace-Beltrami operator on the surface. The derivation of Eq. (3.33) uses our earlier result

$$\frac{\delta A}{\delta f} = -2H, \quad (3.34)$$

and the fact that the volume integral may be rewritten as³

$$V = \int dV = \int dA \frac{1}{3} \mathbf{F} \cdot \mathbf{N}, \quad (3.35)$$

$$dV = \frac{1}{3} h dA \text{ for a cone or pyramid of height } h = \mathbf{F} \cdot \mathbf{N}$$

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$$\Rightarrow \frac{\delta V}{\delta f} = 1.$$

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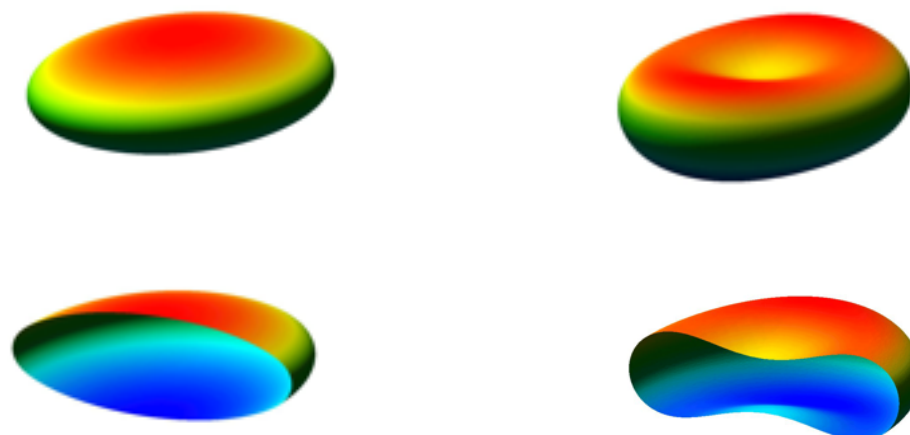
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For **open** membranes with boundary ∂M , a plausible energy functional is given by

$$F_o = \int dA f_c + \sigma \int dA + \gamma \oint_{\partial M} ds, \quad (3.37)$$

where γ is the line tension of the boundary. In this case, variation yields not only the corresponding shape equation but also a non-trivial set of boundary conditions.